

Supplementary Information: Alchemical free energy calculations for nucleotide mutations in protein-DNA complexes

Vytautas Gapsys* and Bert L. de Groot*

Computational Biomolecular Dynamics Group, Max Planck Institute for Biophysical Chemistry, Am Fassberg 11, 37077 Göttingen, Germany

E-mail: vgapsys@gwdg.de; bgroot@gwdg.de

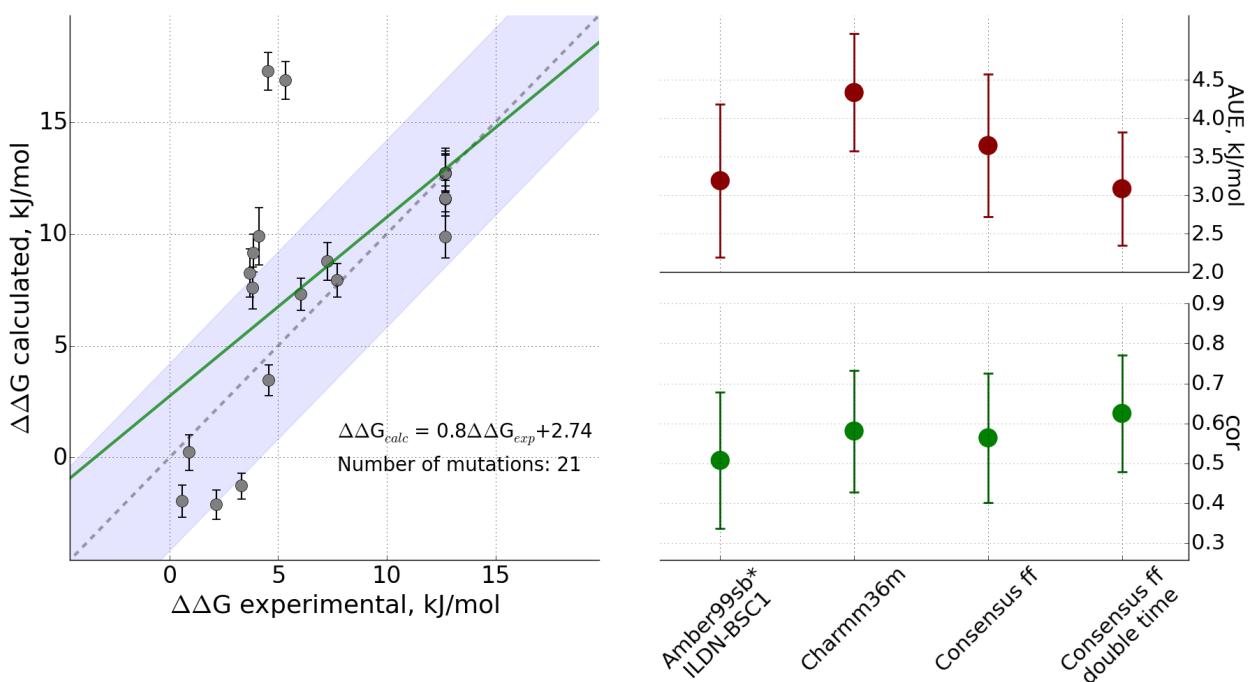
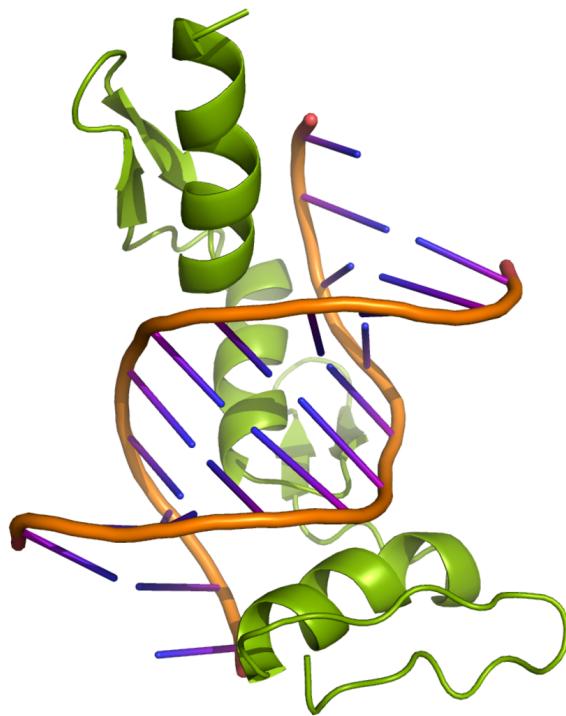


Figure S1: The structure and the calculation results for the DNA bound to the zinc finger protein Zif268. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

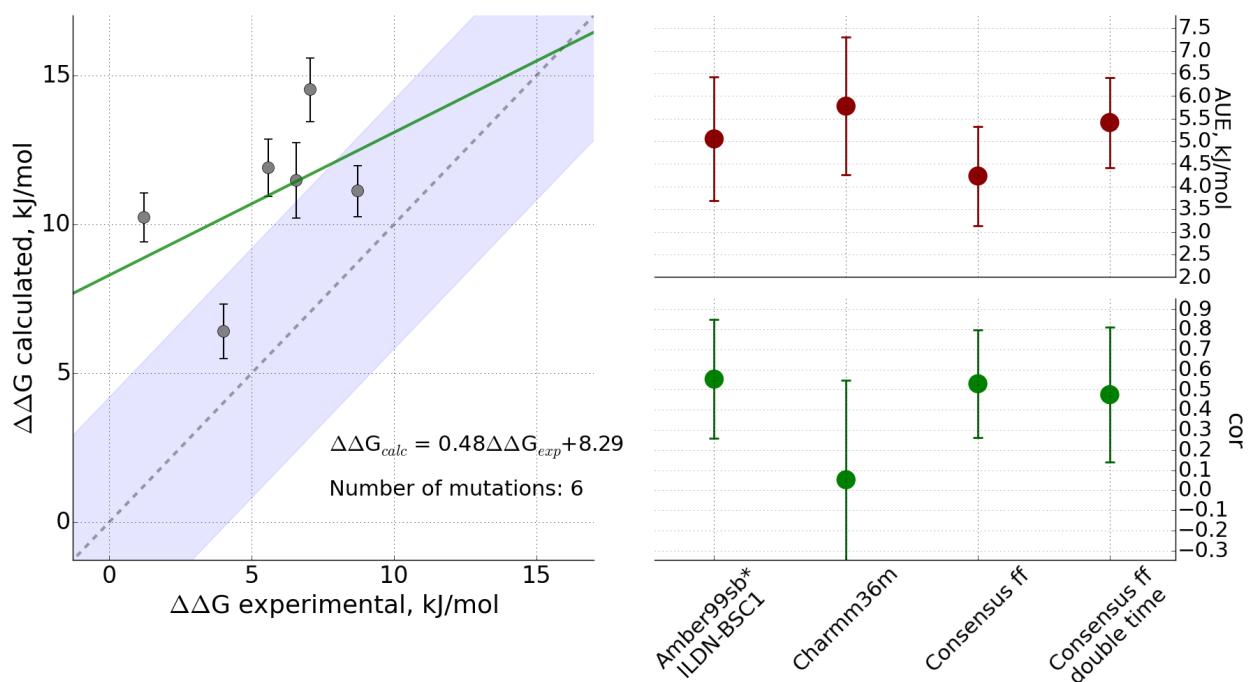
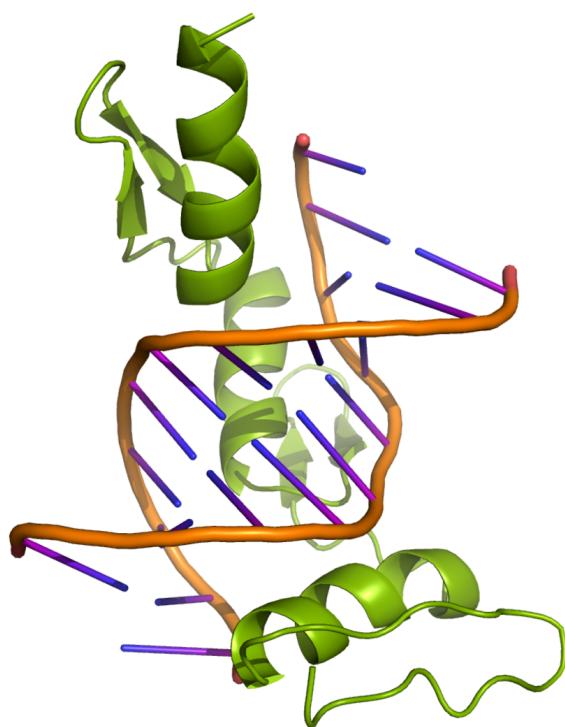


Figure S2: The structure and the calculation results for the DNA bound to the zinc finger protein Zif268 D20A mutant. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

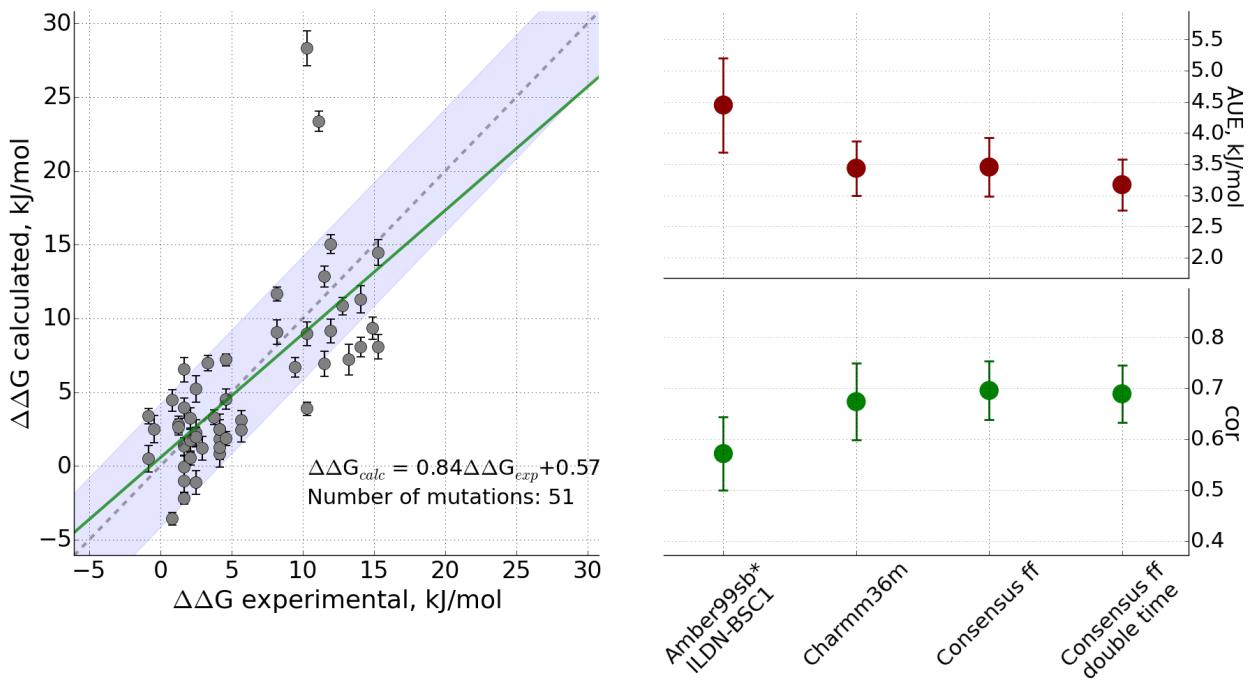
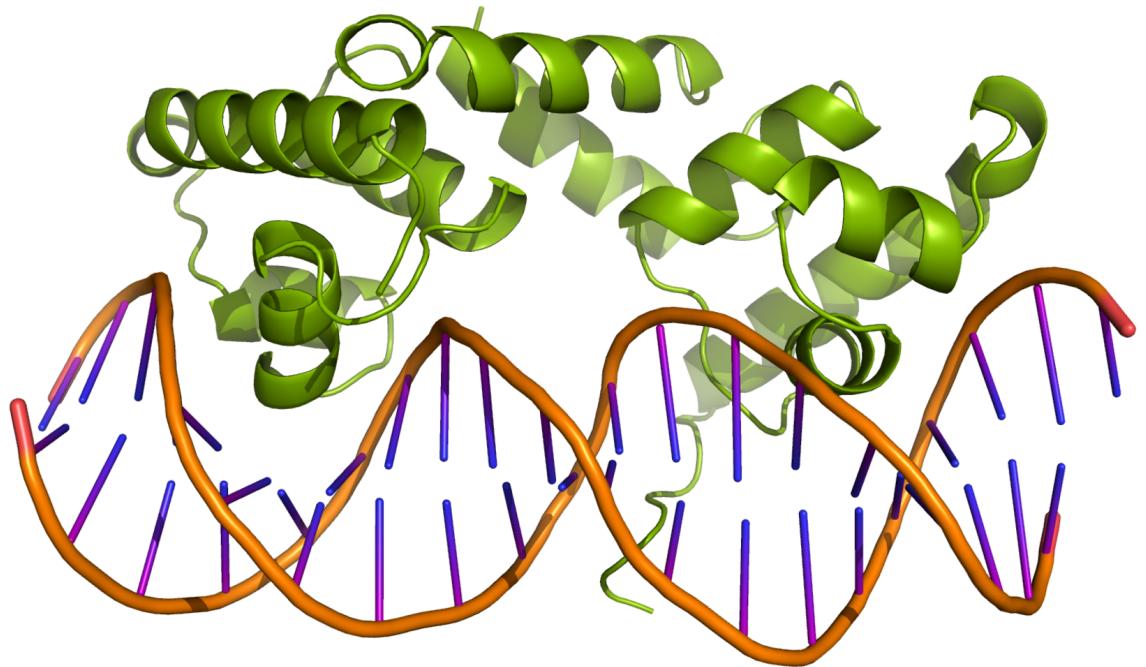


Figure S3: The structure and the calculation results for the DNA bound to the λ repressor λR . Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

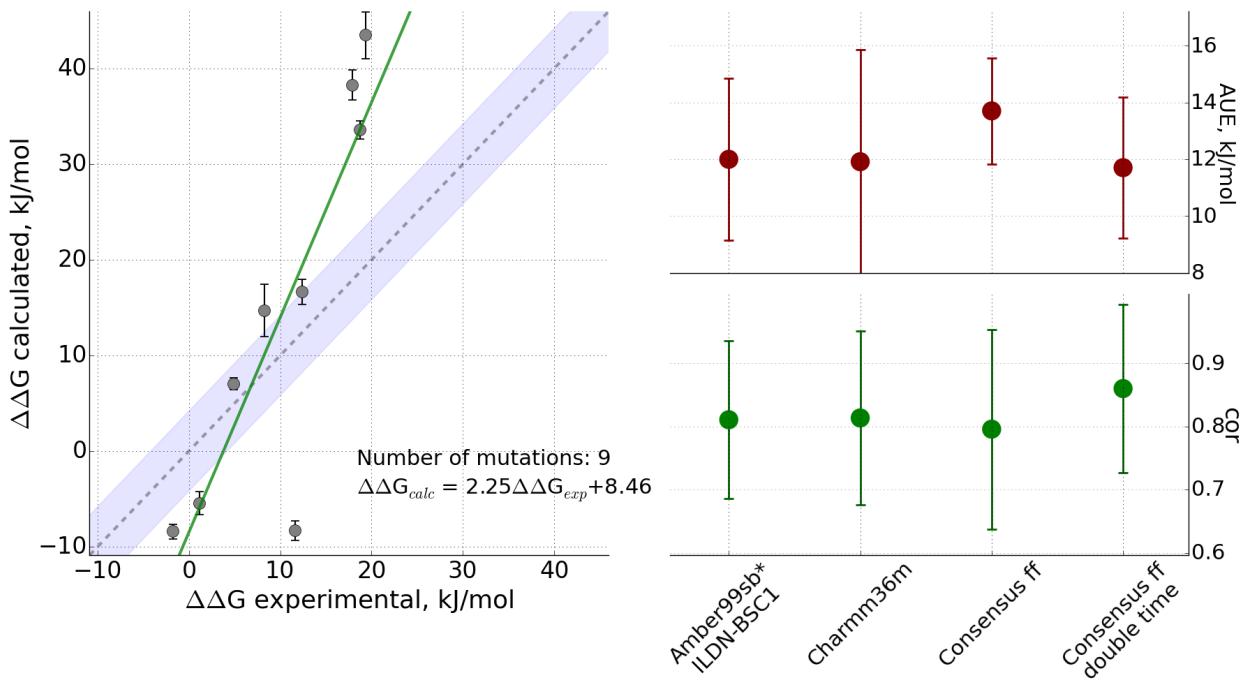


Figure S4: The structure and the calculation results for the DNA bound to the Trp repressor TrpR. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

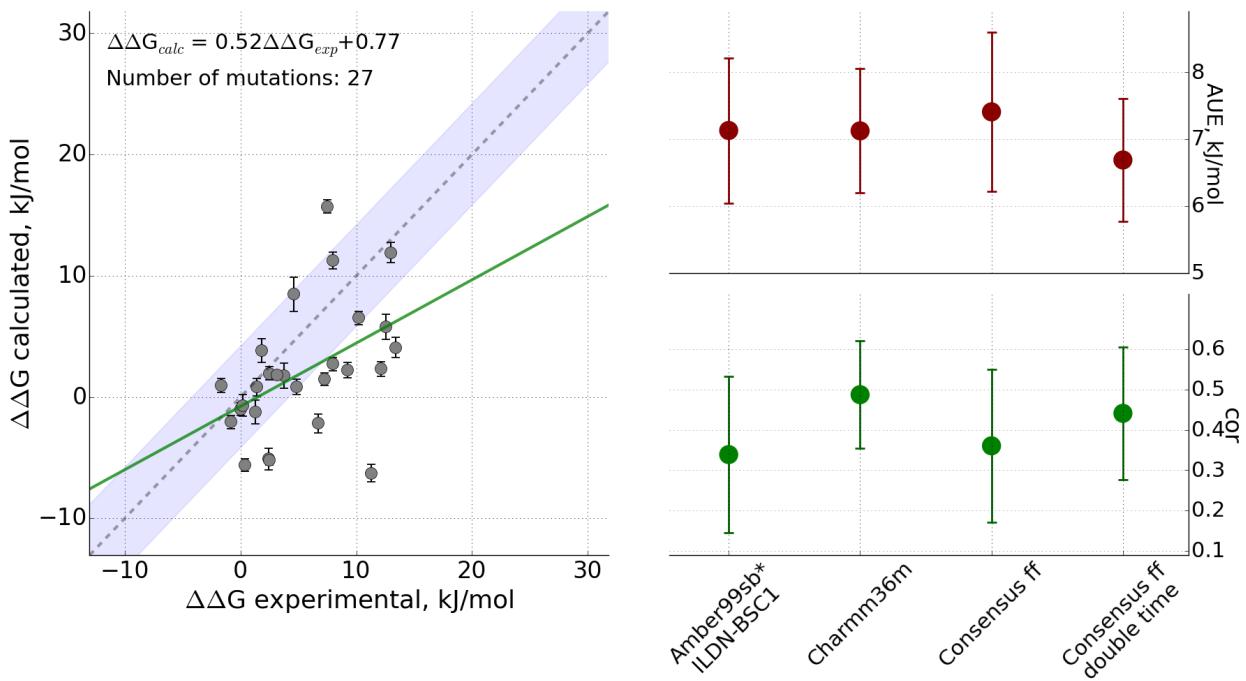
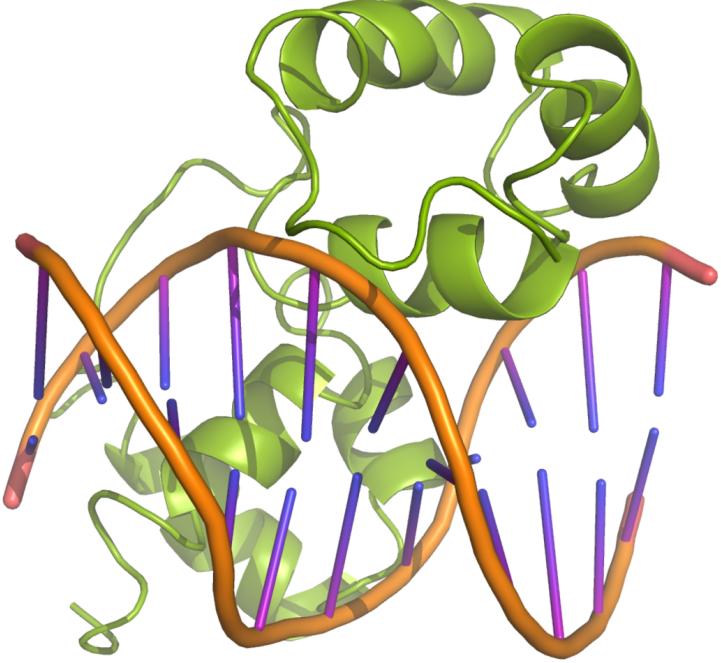


Figure S5: The structure and the calculation results for the DNA bound to the protooncogene c-Myb. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

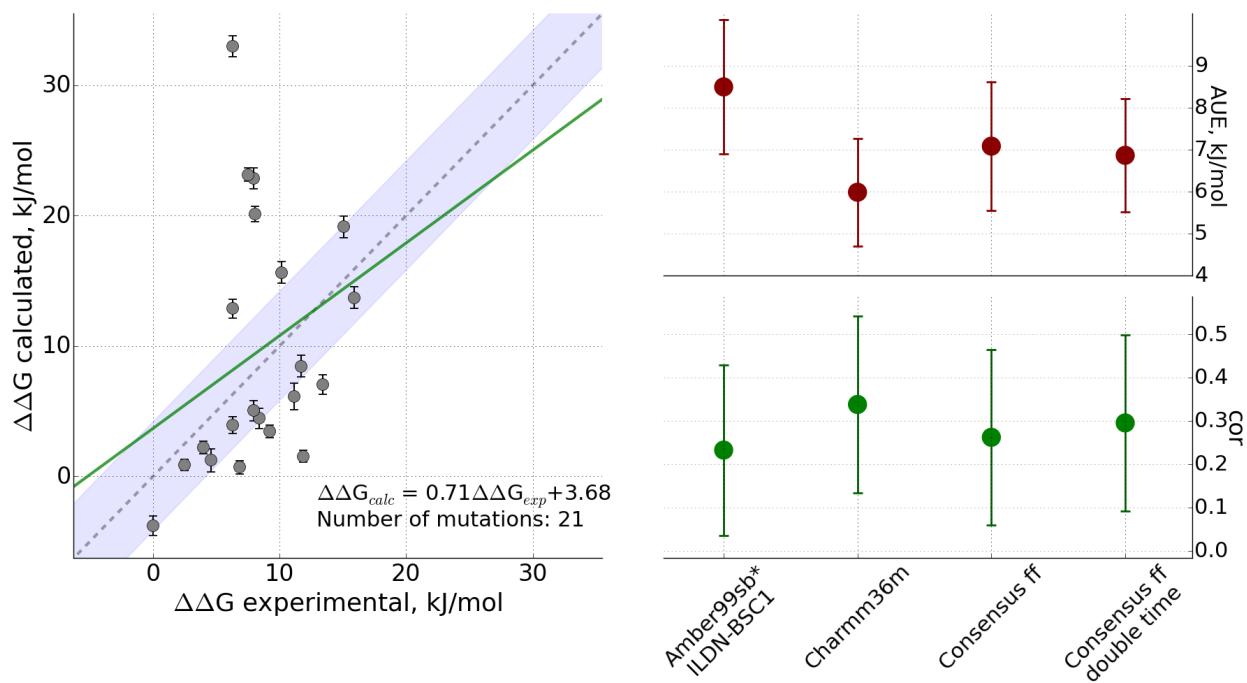
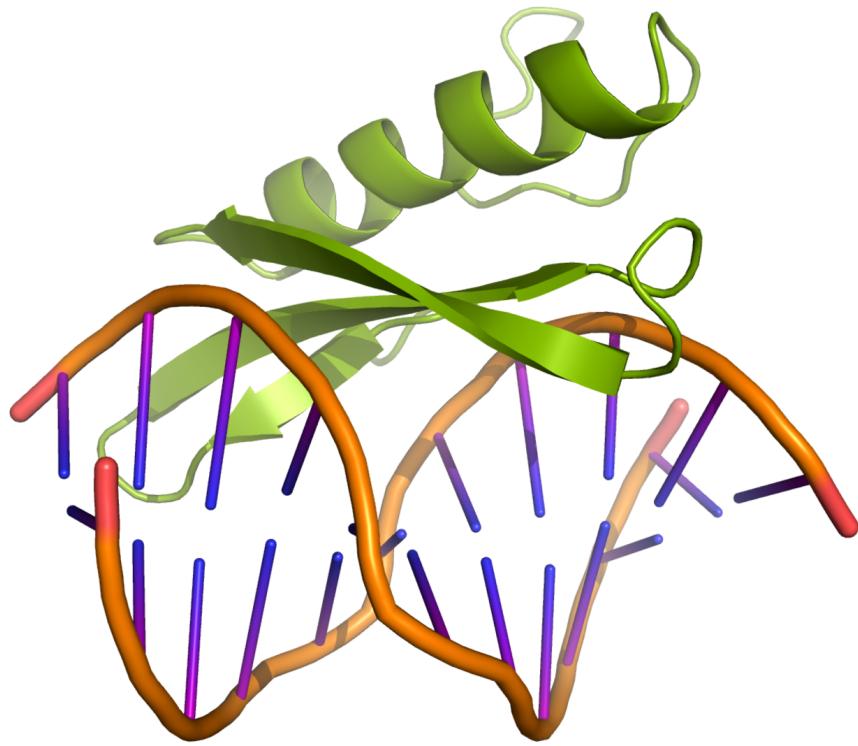


Figure S6: AtERF1. The structure and the calculation results for the DNA bound to the ethylene responsive transcription factor AtERF1. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

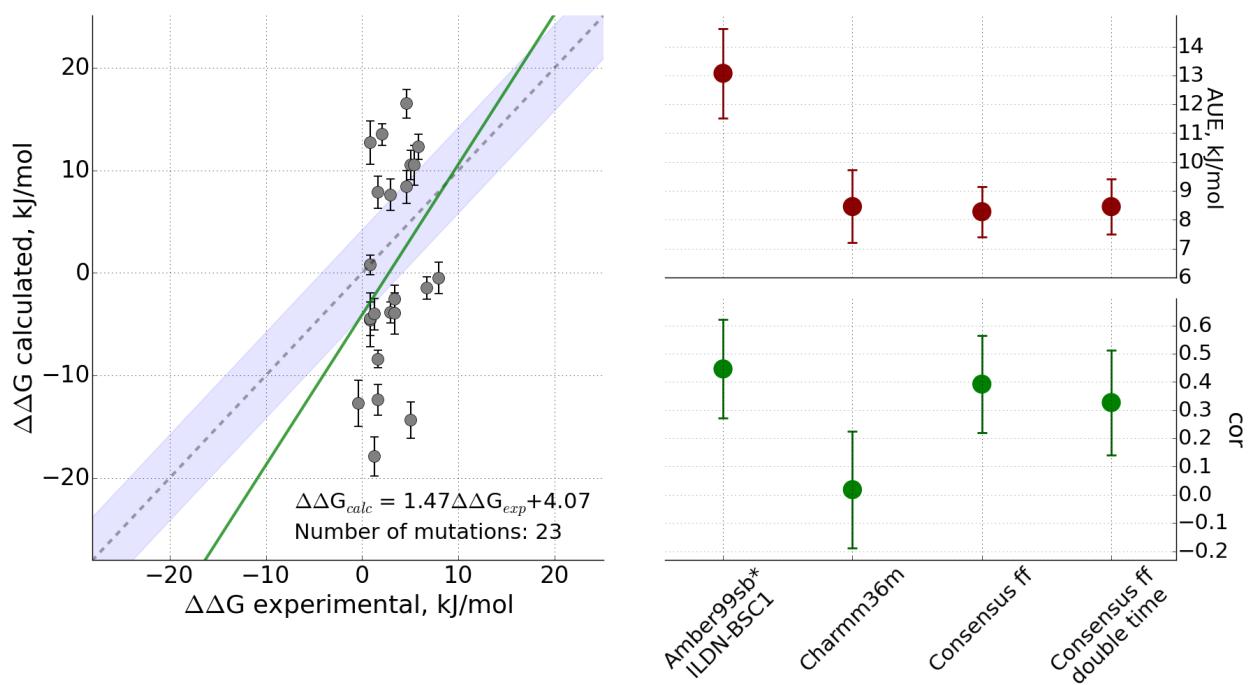
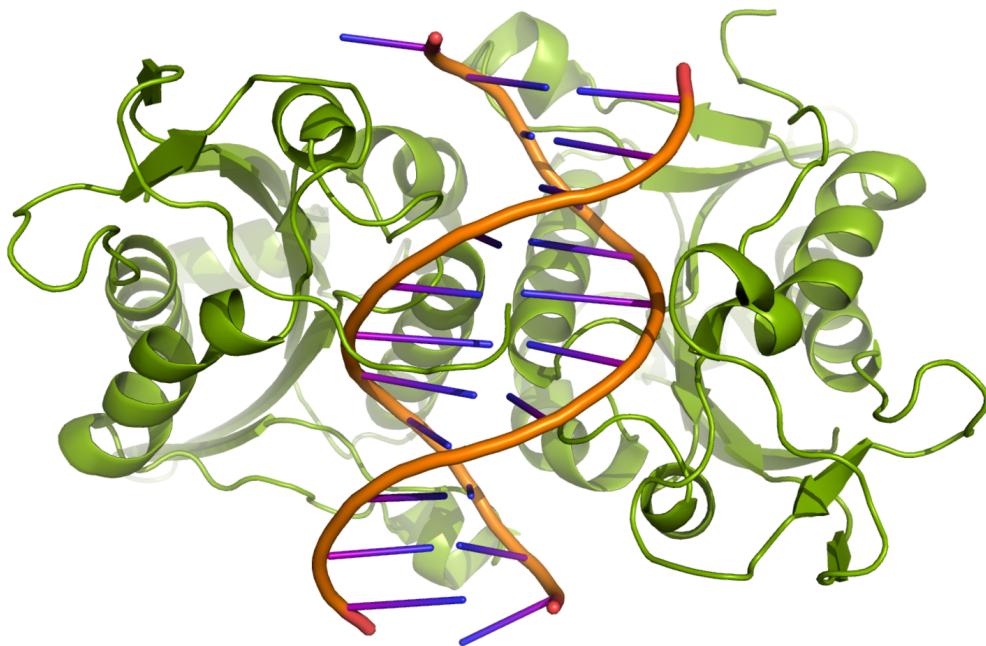


Figure S7: The structure and the calculation results for the DNA bound to the endonuclease BamHI. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

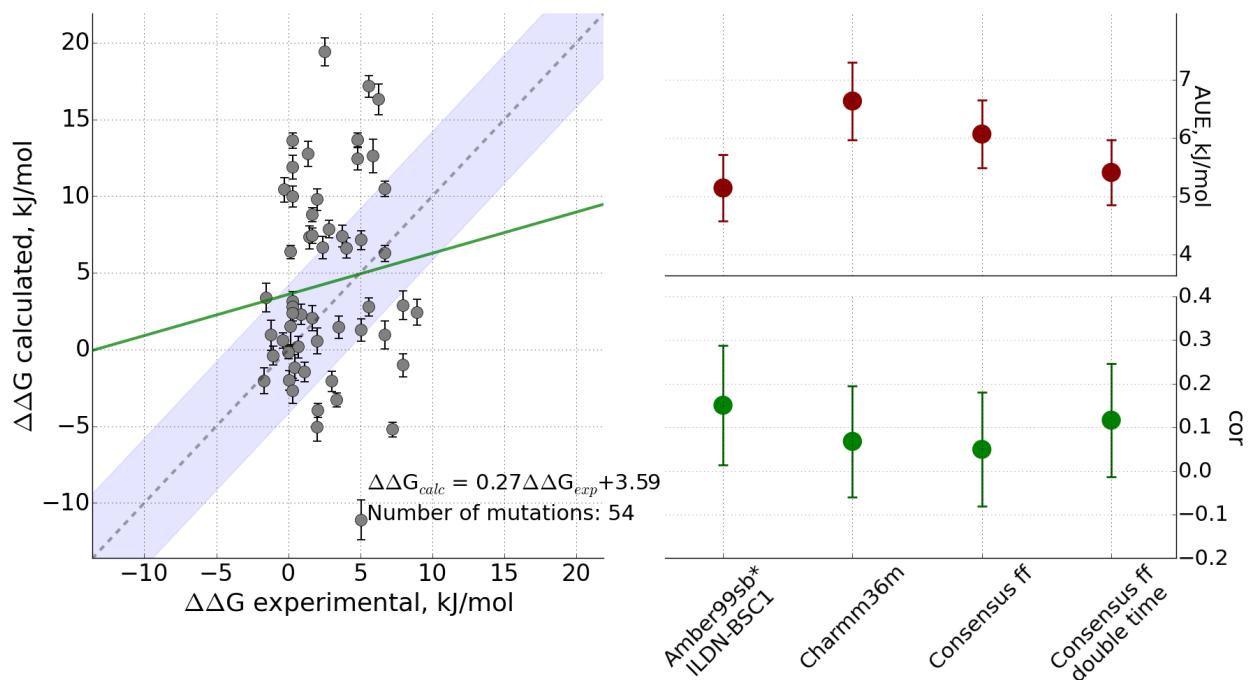
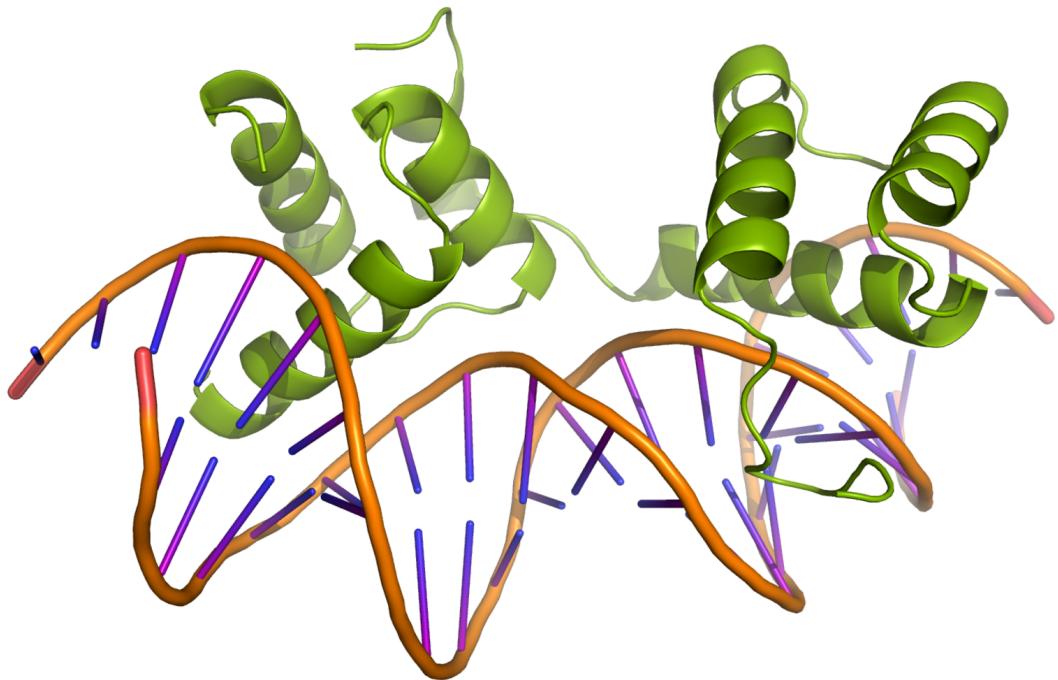


Figure S8: The structure and the calculation results for the DNA bound to the MAT $\alpha 1/\alpha 2$ homeodomain. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

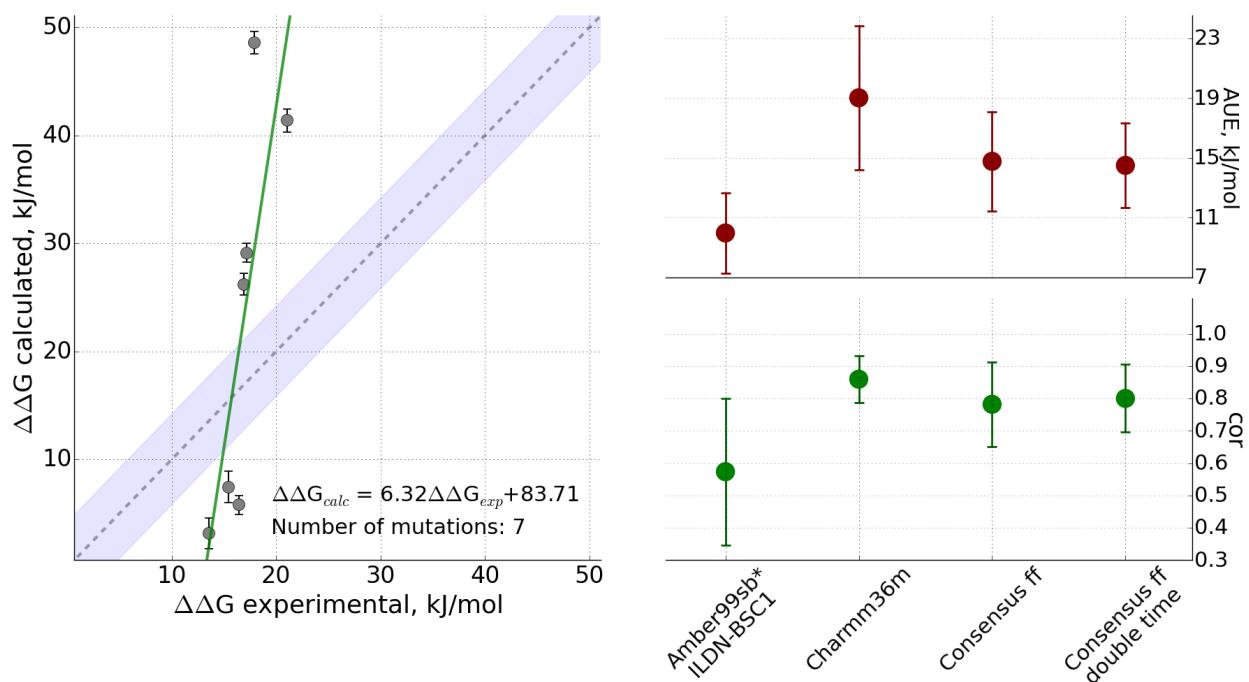
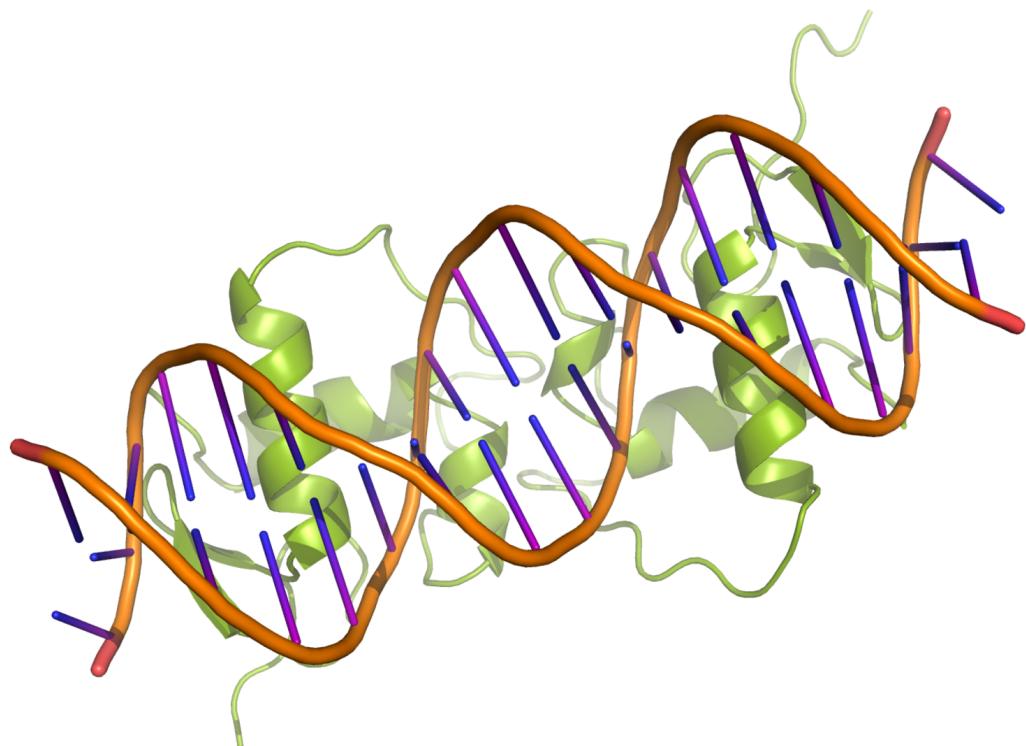


Figure S9: The structure and the calculation results for the DNA bound to the estrogen receptor α (ER). Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

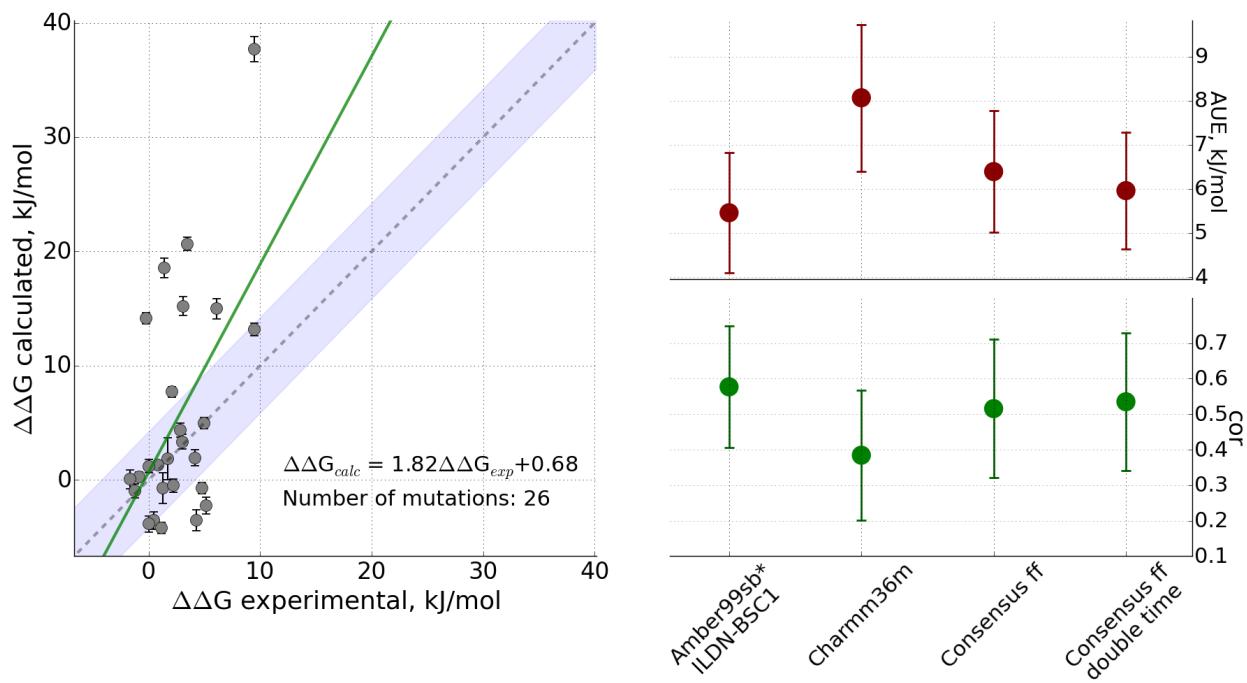
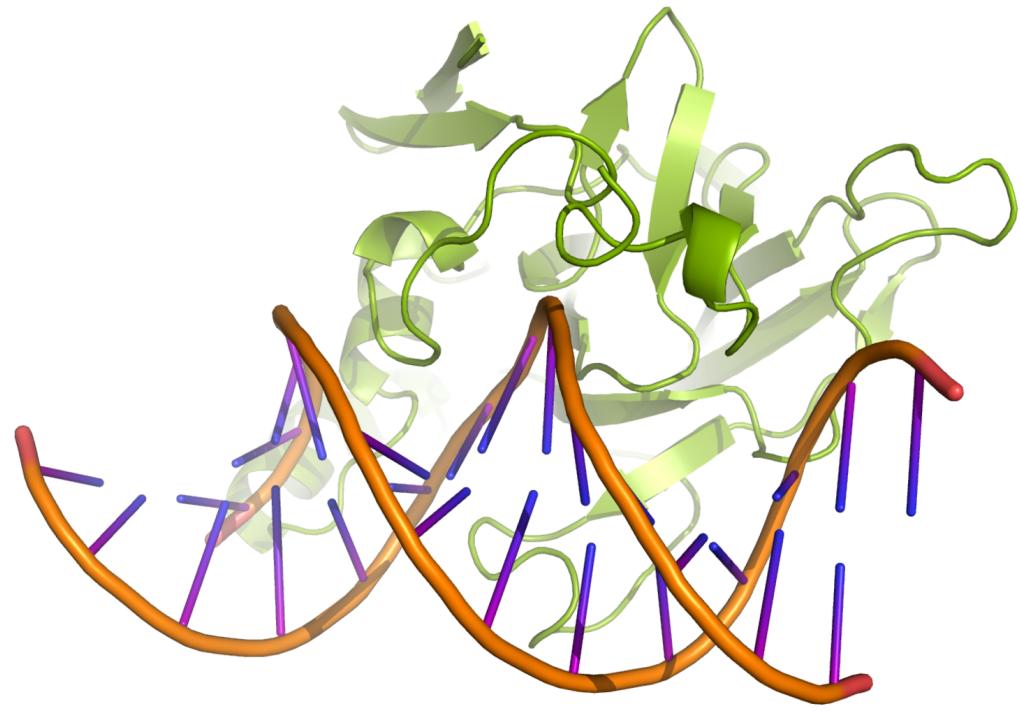


Figure S10: The structure and the calculation results for the DNA bound to the transcriptional activator Ndt80. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

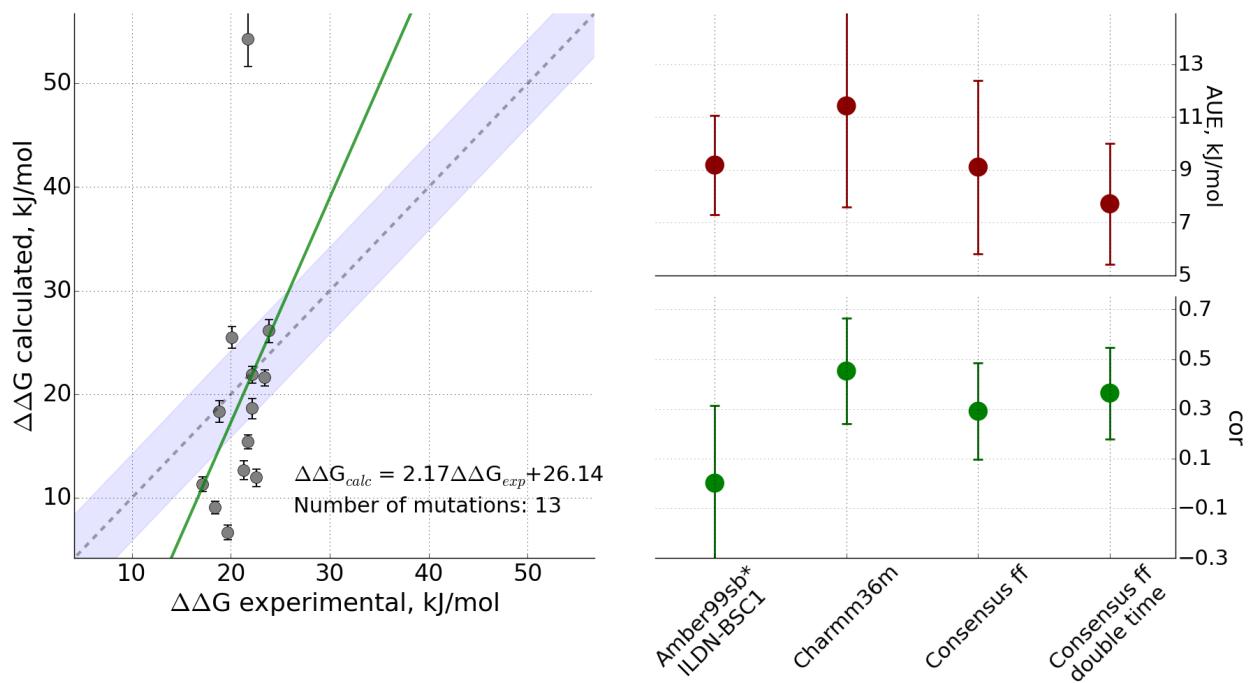


Figure S11: The structure and the calculation results for the DNA bound to the endonuclease EcoRI. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

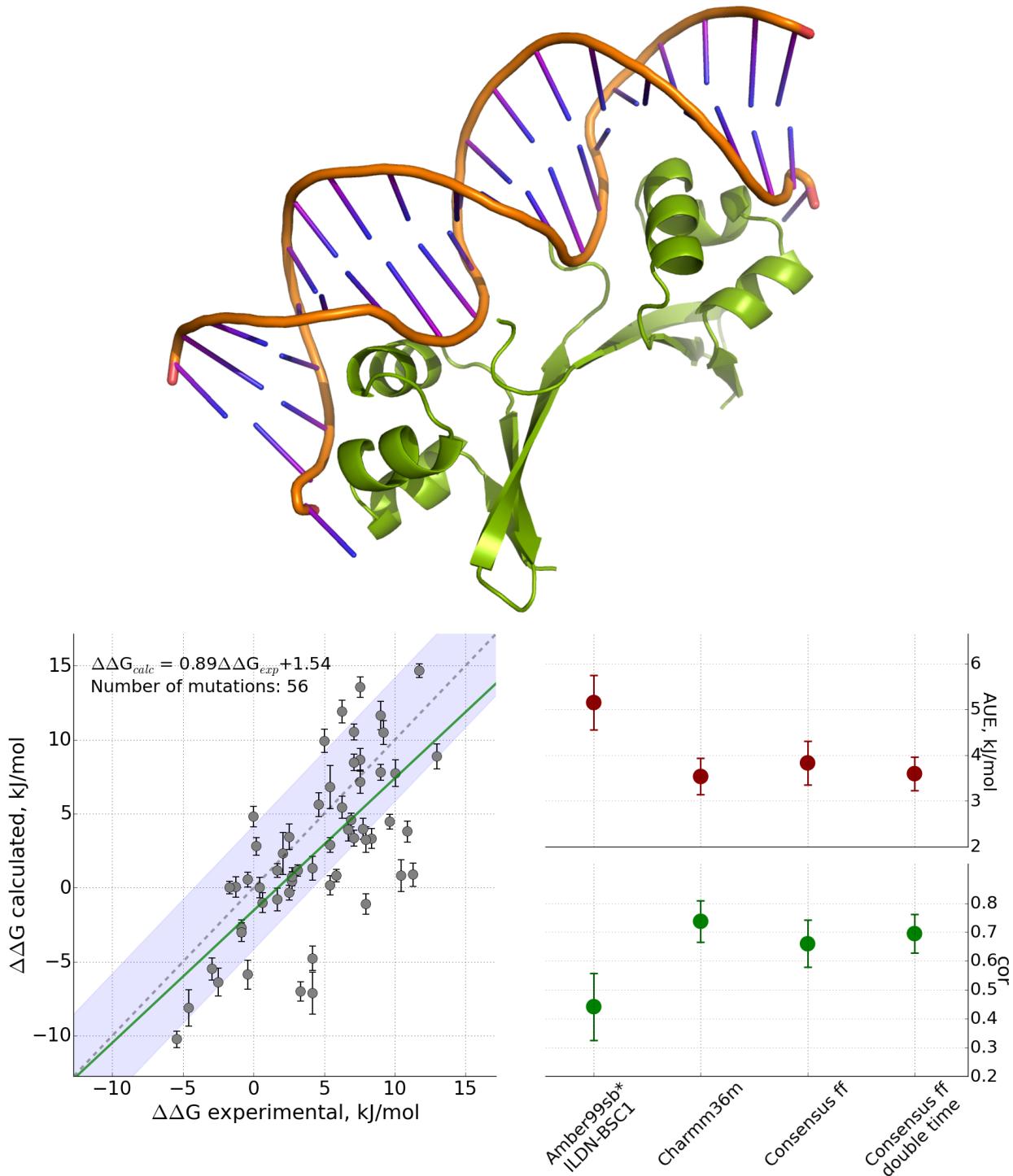


Figure S12: The structure and the calculation results for the DNA bound to the CRO repressor CroR. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

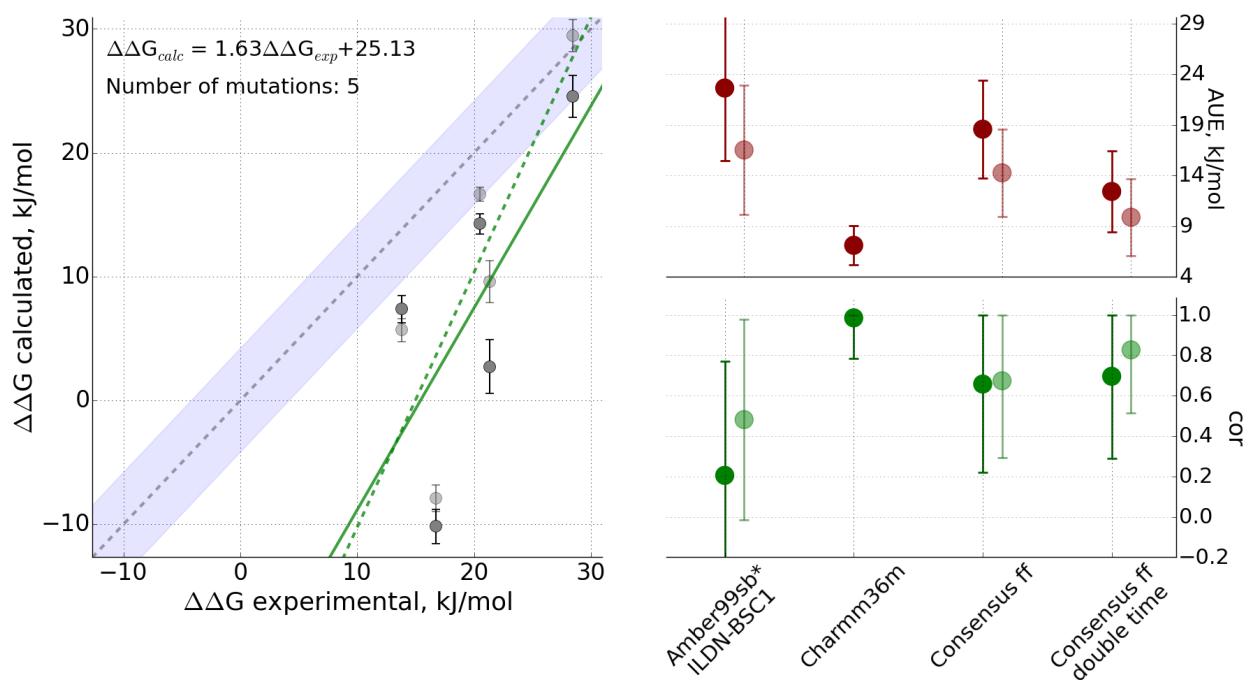
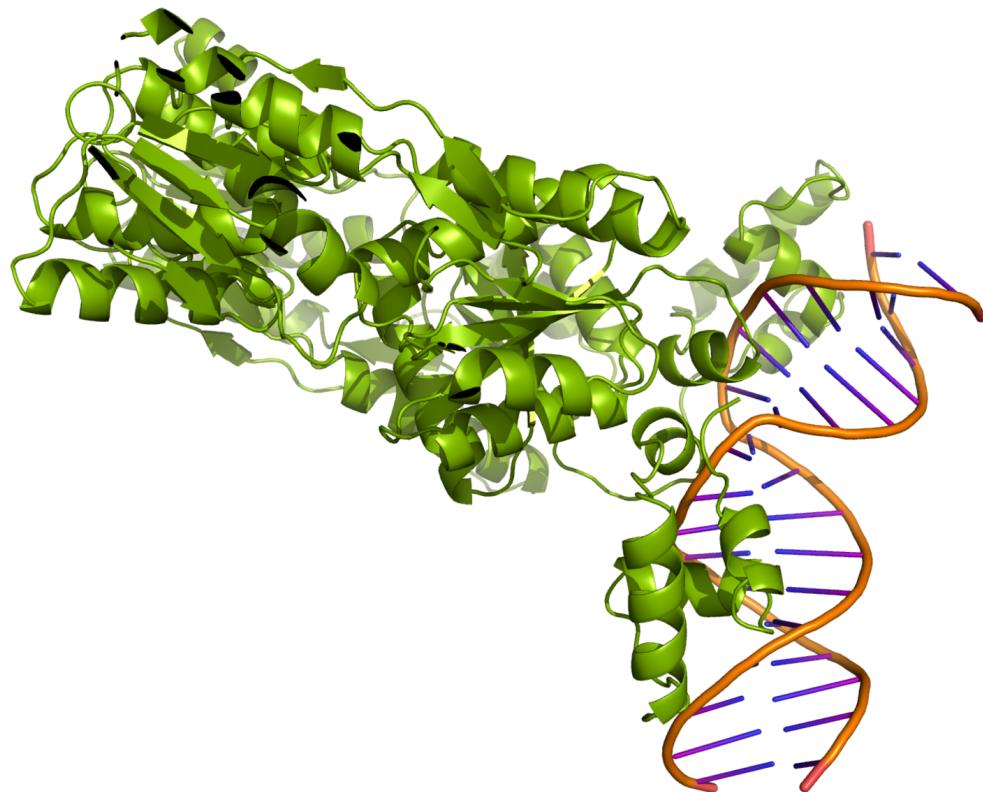


Figure S13: The structure and the calculation results for the DNA bound to the *Lac* repressor LacR. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

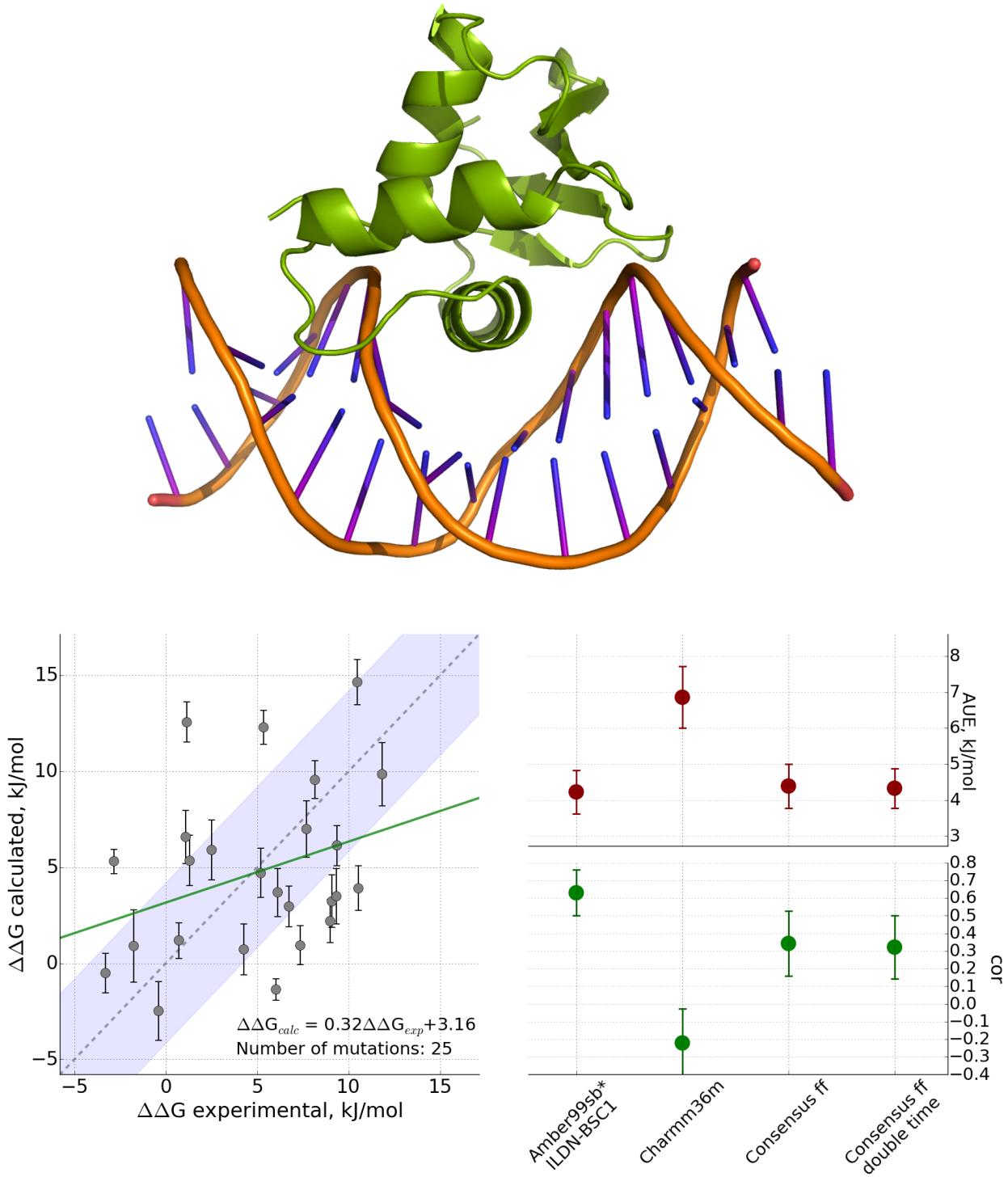


Figure S14: The structure and the calculation results for the DNA bound to the ETS domain of the transcription factor PU.1. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

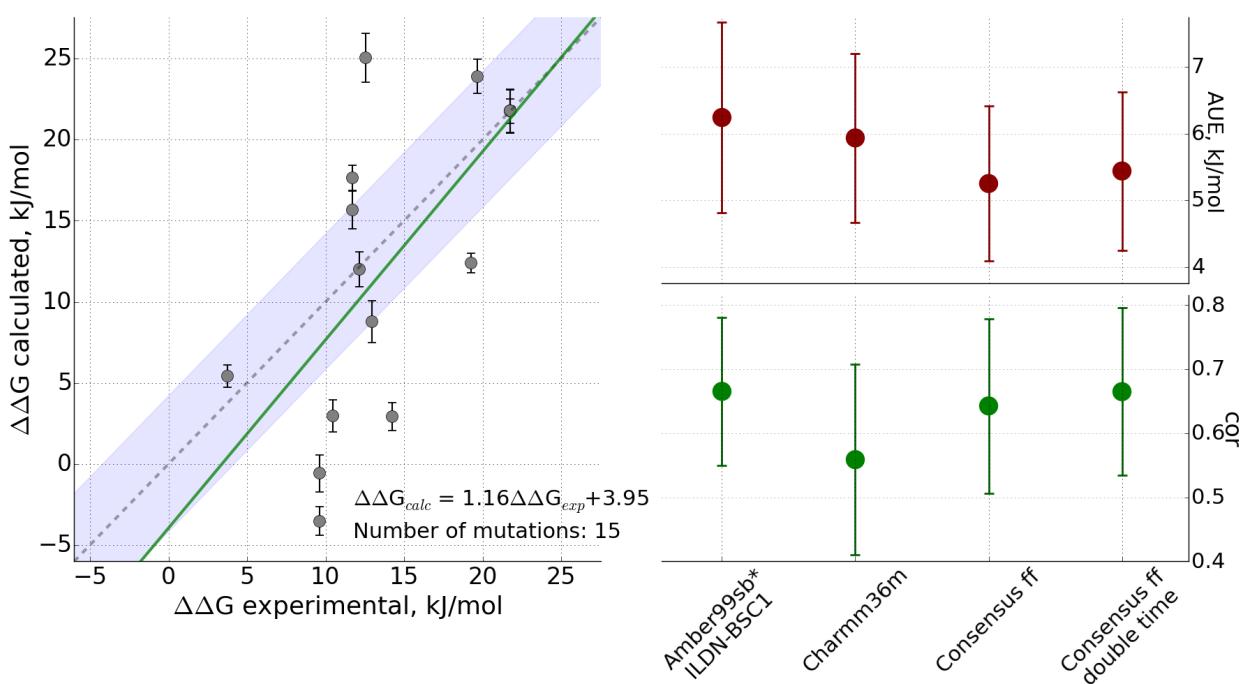
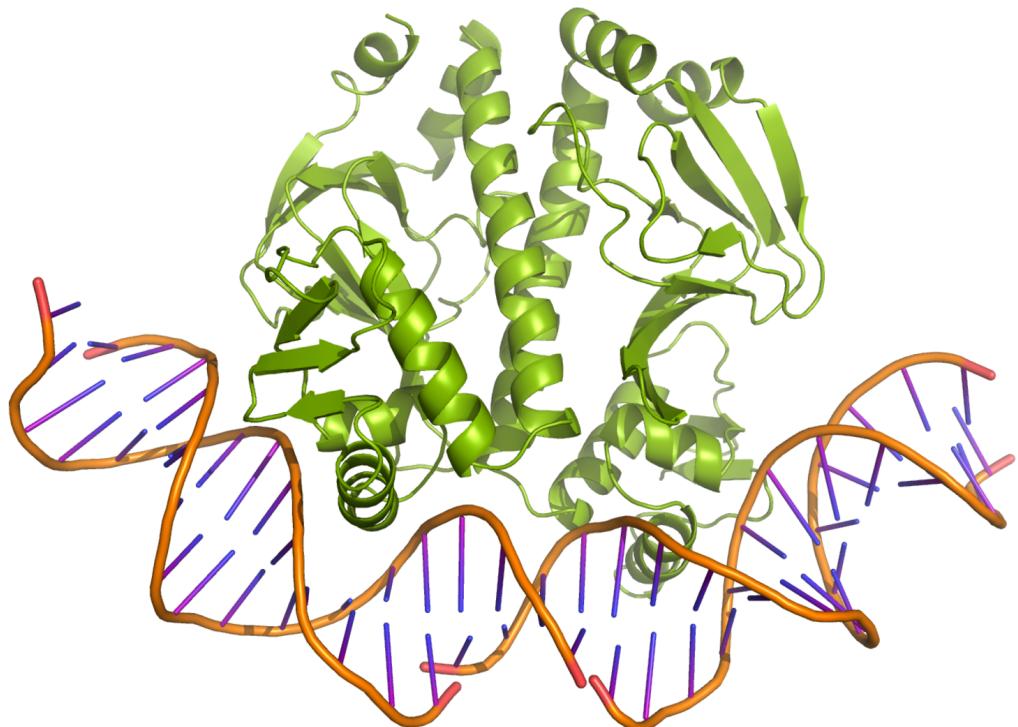


Figure S15: The structure and the calculation results for the DNA bound to the catabolite gene activator protein CAP. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

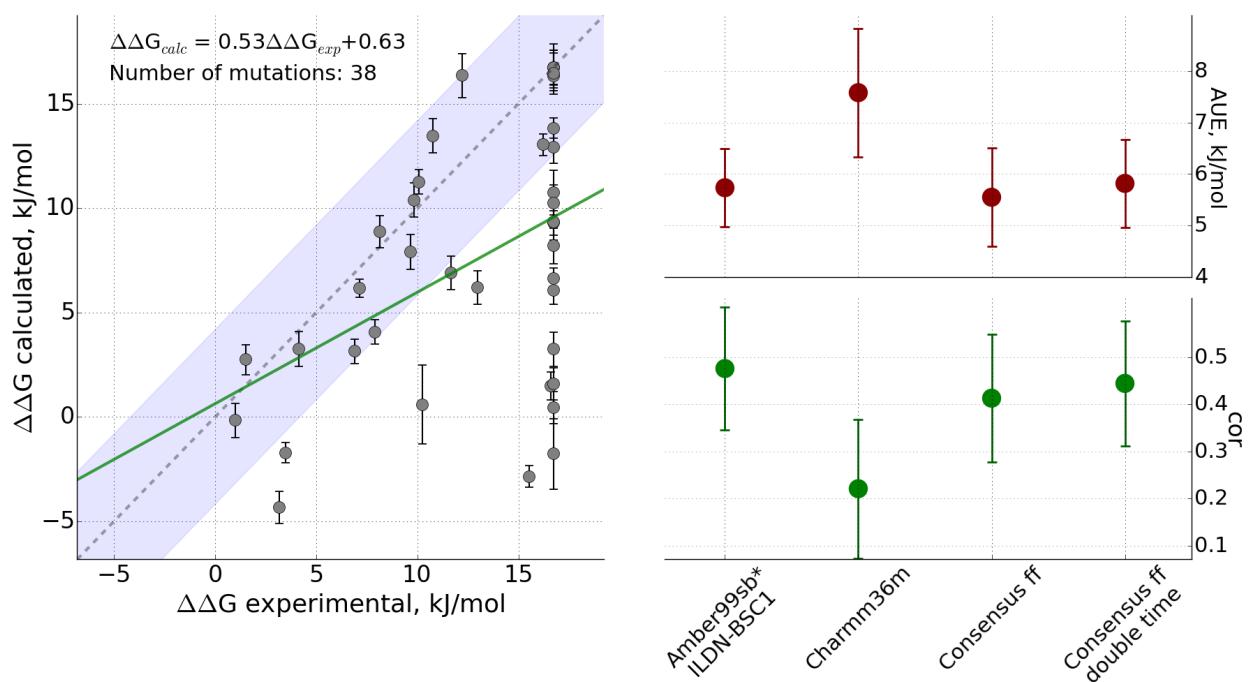
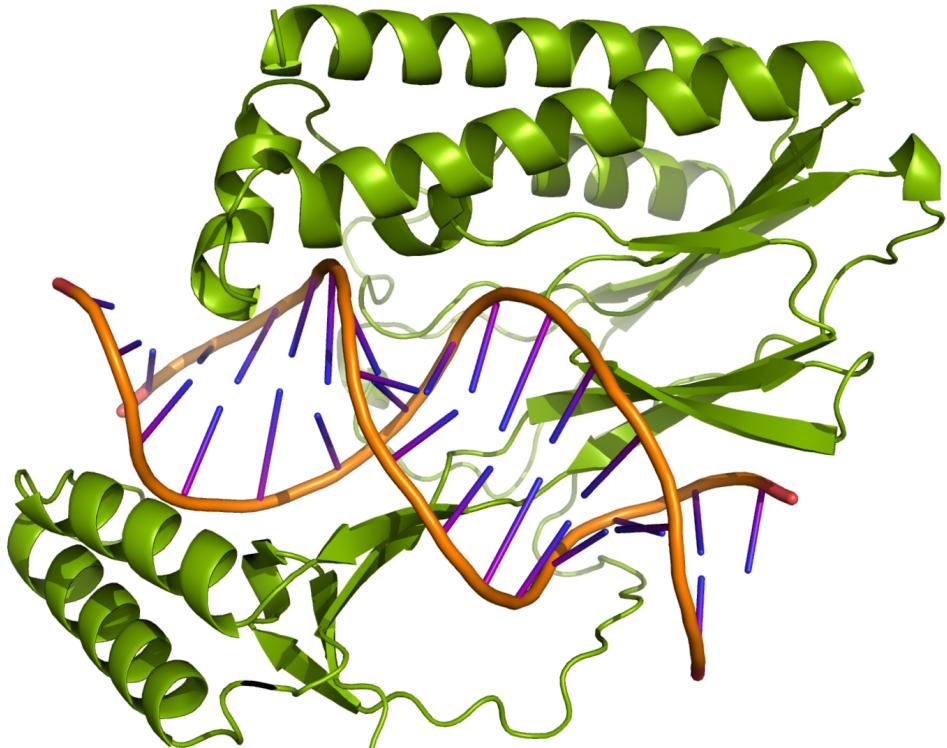


Figure S16: The structure and the calculation results for the DNA bound to the Tus protein. Panel on the left shows the experimental $\Delta\Delta G$ values against the consensus force field (with the doubled simulation time) calculation. Panel on the top-right depicts the average unsigned error (AUE) of the calculations from the experimental results, while panel on the bottom-right shows correlations between the calculations and experiments.

Force field comparison

For a better understanding of the error cancellation effects in the concensus force field approach, we performed a detailed investigation where the discrepancies between Amber99sb*ILDN-BSC1 and Charmm36m occur. For the concensus approach to yield results more accurate than those of the individual force fields taken separately, the errors made by the individual force fields must be pointing in opposite directions. However, this effect is not apparent when all the analyzed data values are pooled together (Fig S17).

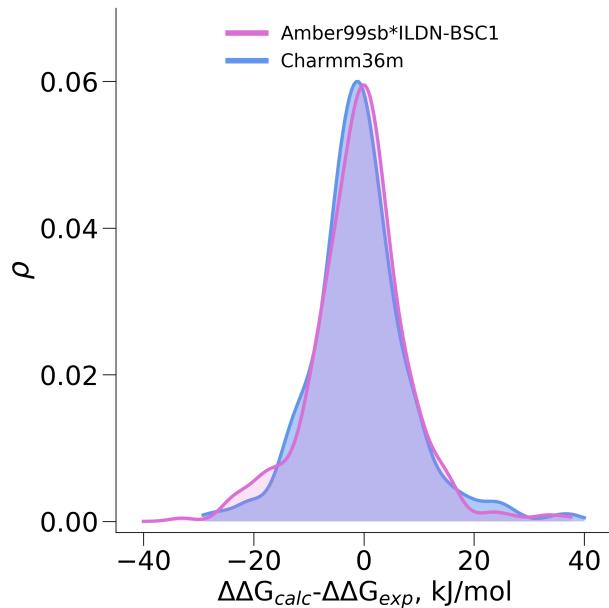


Figure S17: Distributions of the differences between the calculated and experimental $\Delta\Delta G$ values for the Amber99sb*ILDN-BSC1 and Charmm36m force fields.

Clearer trends were revealed by dividing the calculated values into groups, where both force fields were either making an error in the same direction with respect to the experimental $\Delta\Delta G$, or the errors were of opposite signs (Fig S18). From this analysis it is evident that for more than 30% of the investigated mutations the force field results disagree in estimating the $\Delta\Delta G$ values to be smaller or larger than the experimental measurement.

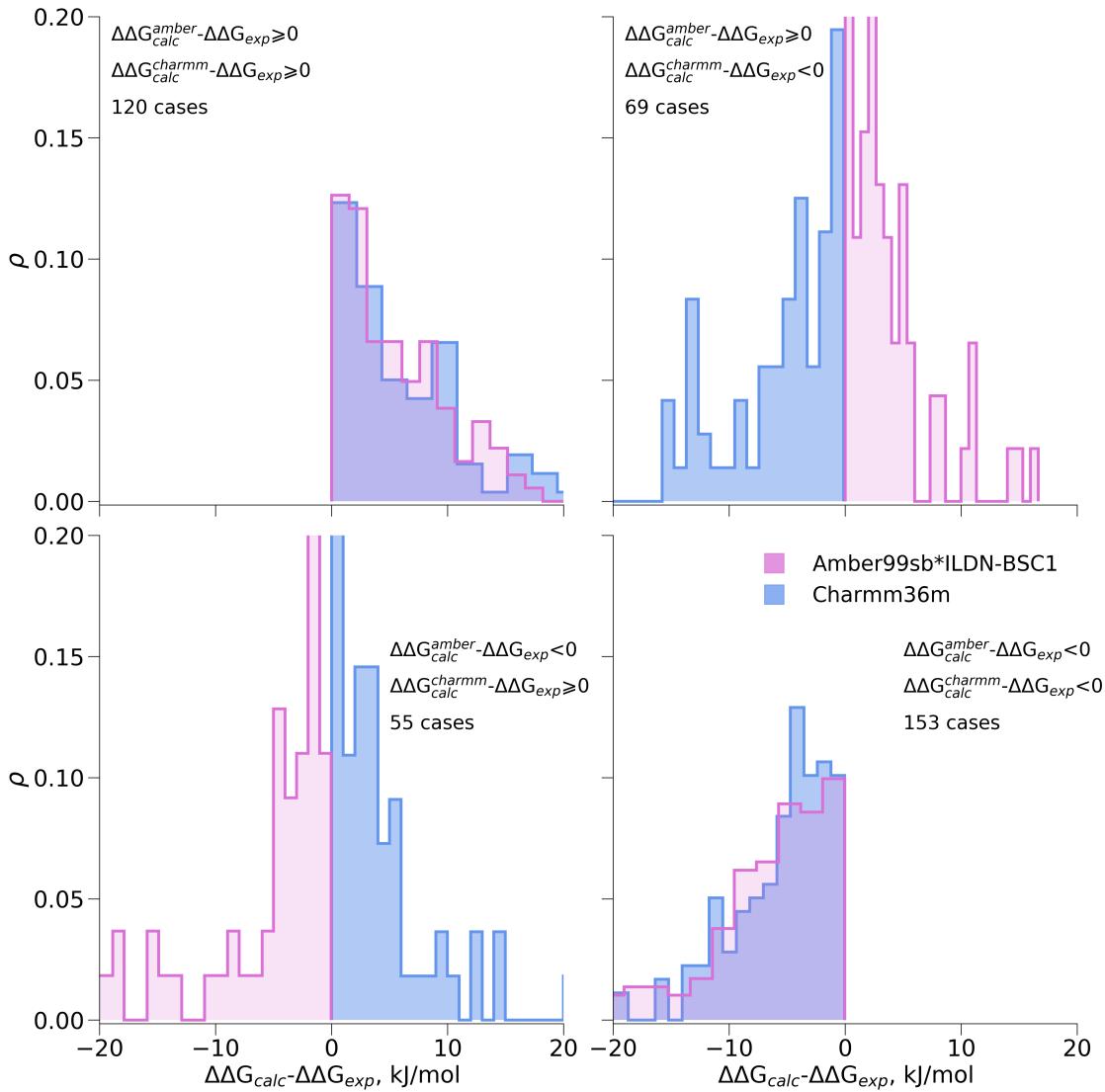


Figure S18: Distributions of the differences between the calculated and experimental $\Delta\Delta G$ values divided into groups by the direction of an error made by individual force fields with respect to the experimental value.

The compensatory effects observed in Fig S18, however, cannot be unambiguously ascribed to the individual protein-DNA complexes (Figs S19). It appears that for most of the systems analysed there is no clear trend for the force fields to yield $\Delta\Delta G$ values that would disagree in terms of a direction from the experimental measurement.

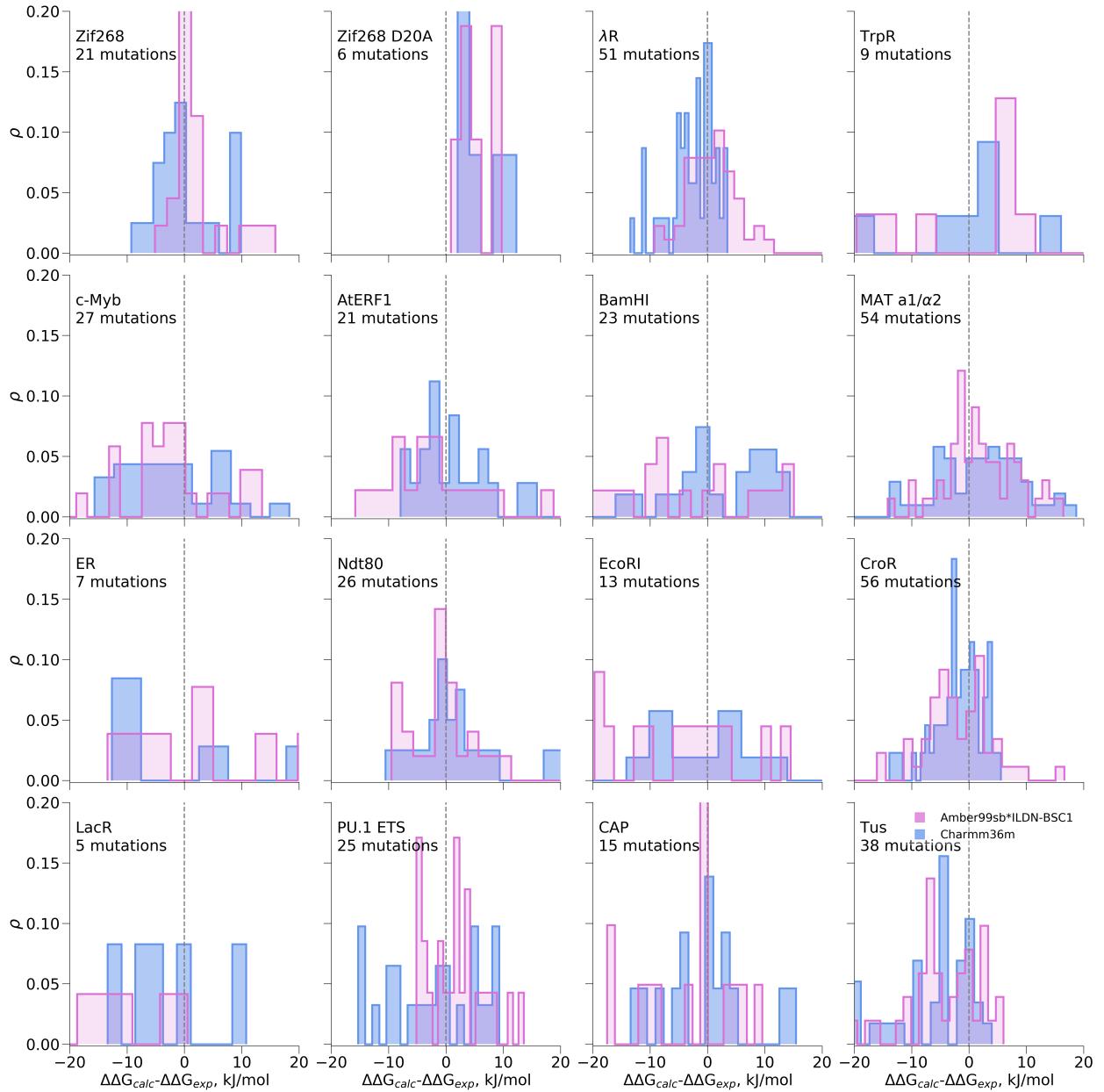


Figure S19: Distributions of the differences between the calculated and experimental $\Delta\Delta G$ values depicted for the individual protein-DNA complexes separately.

While from the distributions of the $\Delta\Delta G_{calc}$ - $\Delta\Delta G_{exp}$ (Fig S19) no clear signal could be discerned, averaging of these differences for every protein-DNA system separately, reveals several cases where the force fields on average point in the opposite directions from the experimentally measured values (Fig S20). Namely, for the λ R and PU.1 ETS complexes, Amber99sb*ILDN-BSC1 on average predicts $\Delta\Delta G$ to be too high, while Charmm36m estimates too low $\Delta\Delta G$ values; the situation is reversed for the BamHI and EcoRI complexes. These systems could potentially be of interest for the further investigation into the differences between the force fields.

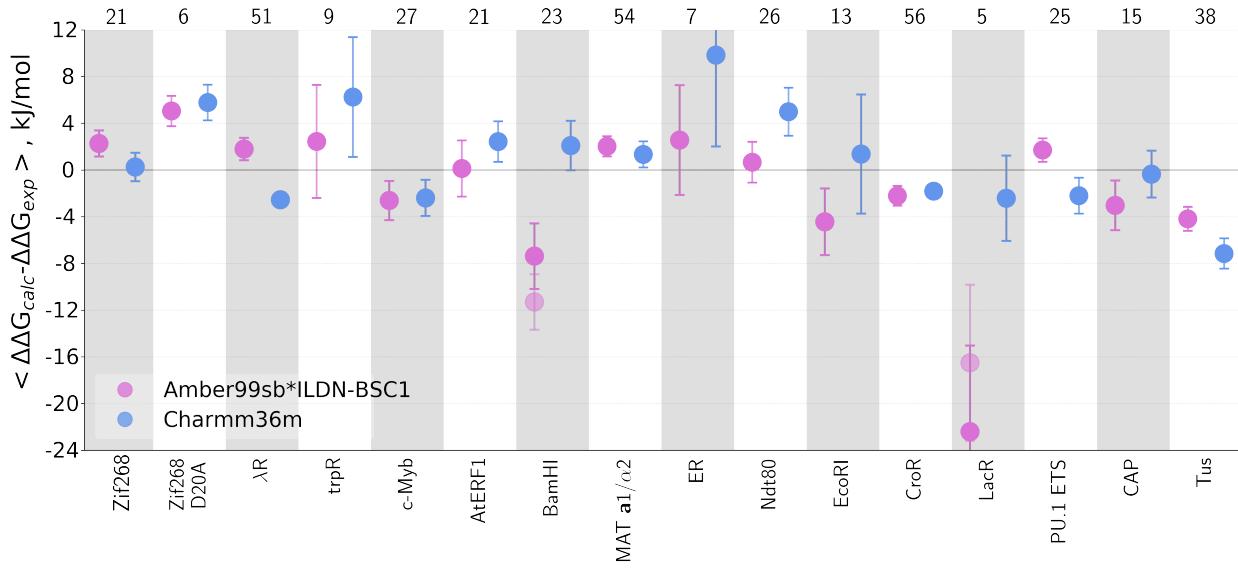


Figure S20: Averages of the absolute differences between the calculated and experimental $\Delta\Delta G$ values for the individual protein-DNA complexes.

Table S1: Experimental and calculated $\Delta\Delta G$ values for the protein-DNA complexes upon nucleotide mutation

Nucleotide sequence	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	Reference
	Amber99sb*ILDN-BSC1	Charmm36m	Experimental	
Zif268				
GCGTGGGC GT				
CCGTGGGC GT	6.64 ± 1.95	13.16 ± 3.50	4.11	1
GAGTGGGC GT	5.76 ± 1.60	1.14 ± 1.11	4.55	1
GGGTGGGC GT	4.80 ± 2.59	13.48 ± 1.42	3.86	1
GTGTGGGC GT	-0.10 ± 0.79	-3.80 ± 0.79	0.57	1
GCGGGGGC GT	6.76 ± 1.74	-6.33 ± 1.58	0.89	1
GCGTGGGAGT	4.59 ± 2.02	9.99 ± 2.69	6.06	1
GCGTGGGGGT	9.43 ± 1.97	6.43 ± 2.49	7.72	1
GCGTGGGTGT	-1.82 ± 1.11	-0.74 ± 0.64	3.33	1
GCCTGGGC GT	10.32 ± 2.68	9.39 ± 2.42	12.71*	1
GCGAGGGC GT	12.71 ± 1.89	12.71 ± 1.56	12.71*	1
GCGCGGGC GT	12.71 ± 0.95	9.84 ± 0.74	12.71*	1
GCGTCGGC GT	12.71 ± 1.80	12.71 ± 2.01	12.71*	1
GCGTGCGC GT	12.71 ± 3.98	12.71 ± 4.59	12.71*	1
GCGTGGCC GT	12.71 ± 3.17	12.71 ± 1.68	12.71*	1
GCGTGGGC CT	12.71 ± 2.46	7.49 ± 1.59	12.71*	1
GCGTGGGC GG				
GCGTGGGAGG	5.63 ± 1.95	9.50 ± 1.62	3.82	2
GCGTGGGGGG	9.79 ± 1.61	7.74 ± 2.39	7.28	2
GCGTGGGTGG	2.92 ± 1.16	-7.12 ± 0.74	2.15	2
GCGTGGGC AG	14.60 ± 1.44	1.87 ± 1.50	3.71	2
GCGTGGGCC G	18.56 ± 2.52	15.18 ± 4.11	5.33	2
GCGTGGGCTG	20.39 ± 2.00	14.15 ± 1.15	4.55	2
Zif268 D20A				
GCGTGGGCC G				
GCGTGGGAGG	14.15 ± 1.81	9.66 ± 1.39	5.59	2
GCGTGGGGGG	11.49 ± 1.76	10.74 ± 1.40	8.72	2
GCGTGGGTGG	7.40 ± 1.11	15.55 ± 1.48	6.56	2
GCGTGGGC AG	6.70 ± 1.16	6.12 ± 1.53	4.03	2
GCGTGGGCC G	16.78 ± 2.07	12.26 ± 1.32	7.06	2
GCGTGGGCTG	6.97 ± 1.17	13.50 ± 1.45	1.22	2

λR

Nucleotide sequence	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	Reference
	Amber99sb*ILDN-BSC1	Charmm36m	Experimental	
ATACCTCTGGCGGTGATAT				
ATACC A CTGGCGGTGATAT	1.98 ± 1.34	-0.98 ± 1.47	-0.84	3
ATACCTCTGGCGGTGAT T	6.26 ± 0.74	-3.59 ± 0.64	1.67	3
ATACCTCTGGCGGTGATT T	10.46 ± 1.29	2.61 ± 1.18	1.67	3
ATACCTCTGGCGGTGAT C T	5.74 ± 1.25	-2.34 ± 0.88	2.09	3
ATACCTCTGGCGGTGAG A T	6.15 ± 2.25	2.91 ± 1.15	4.60	3
ATACCTCTGGCGGTGA A AT	12.14 ± 1.62	13.57 ± 1.52	11.55	3
ATACCTCTGGCGGTGAC A T	14.01 ± 1.22	9.30 ± 0.65	8.20	3
ATACCTCTGGCGGTG G TAT	4.15 ± 1.96	2.62 ± 1.07	-0.84	3
ATACCTCTGGCGGTGTTAT	3.52 ± 2.32	1.49 ± 1.42	-0.42	3
ATACCTCTGGCGGTG C TAT	4.62 ± 5.13	4.28 ± 1.54	0.84	3
ATACCTCTGGCGGT A ATAT	3.85 ± 0.95	1.77 ± 0.92	1.26	3
ATACCTCTGGCGGT T ATAT	6.58 ± 1.92	-4.97 ± 1.86	4.18	3
ATACCTCTGGCGGT C ATAT	0.07 ± 2.57	-2.31 ± 2.12	2.51	3
ATACCTCTGGCGGG G ATAT	17.26 ± 1.52	1.07 ± 2.03	11.97	3
ATACCTCTGGCGG A GATAT	10.51 ± 1.34	7.63 ± 2.12	8.20	3
ATACCTCTGGCGG C GATAT	8.43 ± 0.85	-0.67 ± 0.92	10.29	3
ATACCTCTGGCG A TGATAT	15.59 ± 1.16	6.09 ± 0.69	12.80	3
ATACCTCTGGCG T TGATAT	15.05 ± 1.76	13.88 ± 1.77	15.31	3
ATACCTCTGGCG C TGATAT	12.03 ± 1.85	4.14 ± 4.99	15.31	3
ATACCTCTGGC A GTGATAT	20.48 ± 1.08	9.58 ± 1.71	11.97	3
ATACCTCTGG C TGTGATAT	3.95 ± 1.88	10.50 ± 2.11	13.22	3
ATACCTCTGCC C GTGATAT	11.90 ± 1.32	10.69 ± 1.05	14.06	3
ATACCTCTGG G GGGTGATAT	44.52 ± 1.77	12.11 ± 2.30	10.29	3
ATACCTCTGG A GGGTGATAT	21.08 ± 2.59	-3.18 ± 1.82	10.29	3
ATACCTCTGG T GGGTGATAT	32.71 ± 1.55	14.02 ± 2.28	11.13	3
ATACCTCTG A CGGTGATAT	9.40 ± 0.68	4.97 ± 0.69	4.60	3
ATACCTCTG T CGGTGATAT	9.65 ± 1.64	3.73 ± 1.19	9.46	3
ATACCTCTGCCGGT G ATAT	0.20 ± 1.28	5.96 ± 1.08	5.69	3
ATACCTCTCGCGGT G ATAT	-1.83 ± 1.38	1.71 ± 1.17	1.67	3
ATACCTCTTGC C GGTGATAT	4.86 ± 1.32	1.63 ± 1.09	2.09	3
ATACCTCT A GC C GGTGATAT	0.52 ± 0.94	-7.65 ± 0.71	0.84	3
ATACCTCCGGCGGTGATAT	3.76 ± 0.72	1.51 ± 0.67	1.26	3
ATACCT C AGGC C GGTGATAT	3.77 ± 1.97	10.06 ± 0.95	11.55	3
ATACCT C GGCGGTGATAT	1.86 ± 1.21	1.75 ± 1.03	4.18	3
ATACCT T TGGCGGTGATAT	0.50 ± 0.92	0.59 ± 0.63	2.09	3
ATACCT A TGGCGGTGATAT	8.62 ± 2.27	10.06 ± 2.12	14.90	3
ATACCT G TGGCGGTGATAT	9.51 ± 1.34	6.63 ± 0.99	14.06	3
ATACCC C CTGGCGGTGATAT	3.33 ± 0.70	3.28 ± 0.76	3.77	3

Nucleotide sequence	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	Reference
	Amber99sb*ILDN-BSC1	Charmm36m	Experimental	
ATACCGCTGGCGGTGATAT	0.41 ± 1.71	-2.49 ± 1.34	1.67	3
ATACTTCTGGCGGTGATAT	4.06 ± 1.42	0.92 ± 1.05	4.18	3
ATACATCTGGCGGTGATAT	2.03 ± 1.54	2.95 ± 1.66	4.18	3
ATACGTCTGGCGGTGATAT	5.38 ± 1.46	5.07 ± 1.63	2.51	3
ATATCTCTGGCGGTGATAT	10.85 ± 1.13	3.09 ± 1.17	3.35	3
ATAACTCTGGCGGTGATAT	5.83 ± 1.89	-1.31 ± 1.20	2.51	3
ATAGCTCTGGCGGTGATAT	3.04 ± 1.25	-0.63 ± 1.63	2.93	3
ATCCCTCTGGCGGTGATAT	1.71 ± 1.18	0.83 ± 0.91	4.18	3
ATT CCTCTGGCGGTGATAT	4.28 ± 1.74	0.60 ± 3.22	5.69	3
ATGCCTCTGGCGGTGATAT	0.62 ± 0.93	3.15 ± 0.68	4.60	3
ACACCTCTGGCGGTGATAT	-1.62 ± 0.99	-2.80 ± 0.68	1.67	3
AAACCTCTGGCGGTGATAT	5.72 ± 1.12	2.14 ± 1.06	1.67	3
AGACCTCTGGCGGTGATAT	0.93 ± 1.44	2.99 ± 1.05	2.51	3
trpR				
GAACTAGTTAACTAGTAC				
GAACTAGTTAAGAAGTAC	-13.82 ± 3.24	2.89 ± 1.72	1.13	4
GAACCTCTTAACTAGTAC	-8.04 ± 2.59	-8.61 ± 1.67	11.59	4
GTACTAGTTAACTAGTAC	-10.25 ± 1.34	-6.62 ± 1.45	-1.80	4
GAACTAGTTAACTAGTTC	9.66 ± 2.06	4.39 ± 1.14	4.90	4
CAACTAGTTAACTAGTAG	17.52 ± 4.29	11.86 ± 7.30	8.24	4
GAACTAGAATTCTAGTAC	18.52 ± 9.62	14.75 ± 3.40	12.43	4
GAACGAGTTAACTCGTAC	24.46 ± 1.97	42.74 ± 2.42	18.70	4
GAAAAGAGTTAACTCTTAC	24.56 ± 5.24	52.02 ± 3.17	17.87	4
GCCCCCCTTATCATCATT	51.77 ± 7.27	35.20 ± 6.81	19.37	4
AtERF1				
GAGCCGCCACT				
GCGCCGCCACT	-8.61 ± 1.33	1.03 ± 0.82	0.00	5
GGGCCGCCACT	1.37 ± 0.67	0.40 ± 0.53	2.51	5
GTGCCGCCACT	1.76 ± 1.72	0.73 ± 1.74	4.55	5,6
GAACCGGCCACT	-0.42 ± 0.82	7.30 ± 1.22	9.20	5
GACCCGCCACT	6.76 ± 2.08	10.15 ± 1.43	11.72	5
GATCCGCCACT	2.93 ± 1.46	9.31 ± 1.03	11.15	5,6
GAGACGCCACT	12.57 ± 1.26	13.18 ± 2.43	6.28	5
GAGGCGCCACT	0.91 ± 1.51	7.99 ± 2.90	8.37	5
GAGTCGCCACT	-8.99 ± 1.15	10.38 ± 0.70	6.83	5,6
GAGCAGCCACT	25.19 ± 1.97	20.56 ± 1.21	7.95	5
GAGCGGCCACT	33.68 ± 2.59	32.35 ± 1.51	6.28	5

Nucleotide sequence	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	Reference
	Amber99sb*ILDN-BSC1	Charmm36m	Experimental	
GAGCTGCCACT	16.36 ± 1.64	23.87 ± 1.48	8.09	5,6
GAGCCACCACT	32.01 ± 1.61	14.29 ± 0.96	7.53	5
GAGCCCCCACT	18.70 ± 2.03	19.60 ± 2.70	15.06	5
GAGCCTCCACT	12.40 ± 1.51	18.86 ± 2.91	10.13	5,6
GAGCCGACACT	6.36 ± 1.30	3.71 ± 2.03	7.95	5
GAGCCGGCACT	7.23 ± 2.04	0.63 ± 1.15	6.28	5
GAGCCGTCACT	-0.35 ± 0.71	4.74 ± 0.73	3.96	5,6
GAGCCGCAACT	9.94 ± 2.41	17.48 ± 1.23	15.90	5
GAGCCGCGACT	8.60 ± 1.52	5.46 ± 0.99	13.39	5
GAGCCGCTACT	-0.84 ± 0.78	3.92 ± 0.68	11.86	5,6
MAT a1-α2				
CATGTAATTAAATTACATCA				
AATGTAATTAAATTACATCA	0.43 ± 1.27	-10.54 ± 2.96	2.00	7
GATGTAATTAAATTACATCA	1.94 ± 1.24	-24.16 ± 2.00	5.04	7
TATGTAATTAAATTACATCA	2.31 ± 0.73	-8.89 ± 1.14	3.36	7
CCTGTAATTAAATTACATCA	0.10 ± 2.42	3.99 ± 0.93	1.68	7
CGTGTAATTAAATTACATCA	-1.49 ± 1.37	-2.57 ± 0.81	0.06	7
CTTGTAATTAAATTACATCA	-3.38 ± 1.70	1.04 ± 1.06	0.47	7
CAAGTAATTAAATTACATCA	0.67 ± 1.73	0.44 ± 1.87	2.00	7
CACGTAATTAAATTACATCA	-1.60 ± 1.24	-2.56 ± 0.69	3.02	7
CAGGTAATTAAATTACATCA	2.21 ± 1.97	0.33 ± 0.98	5.04	7
CATAATAATTAAATTACATCA	-6.94 ± 0.94	-3.48 ± 0.92	7.26	7
CATCTAATTAAATTACATCA	-1.35 ± 1.43	-0.70 ± 1.17	7.96	7
CATTTAATTAAATTACATCA	-3.34 ± 1.78	5.21 ± 1.42	6.72	7
CATGAAATTAAATTACATCA	2.69 ± 1.96	3.06 ± 1.33	7.96	7
CATGCAATTAAATTACATCA	6.10 ± 1.17	6.43 ± 0.85	6.72	7
CATGGAATTAAATTACATCA	2.52 ± 1.57	2.34 ± 1.19	8.94	7
CATGTCATTAATTACATCA	5.18 ± 1.59	9.60 ± 1.37	3.75	7
CATGTGATTAATTACATCA	21.24 ± 1.02	6.10 ± 1.34	4.81	7
CATGTTATTAAATTACATCA	-1.52 ± 1.44	1.84 ± 1.47	0.70	7
CATGTACTTAATTACATCA	6.55 ± 1.47	7.72 ± 0.93	5.04	7
CATGTA G TTAATTACATCA	-5.88 ± 1.14	-2.06 ± 0.73	2.07	7
CATGTATTAAATTACATCA	1.71 ± 1.53	4.55 ± 1.38	0.32	7
CATGTAATAATTACATCA	11.93 ± 1.70	8.88 ± 0.85	-0.29	7
CATGTAACTAATTACATCA	4.10 ± 0.95	-4.51 ± 0.56	0.00	7
CATGTAAGTAATTACATCA	3.71 ± 2.01	-0.83 ± 1.13	3.48	7
CATGTAATAAAATTACATCA	10.26 ± 1.71	4.38 ± 1.21	1.45	7
CATGTAATCAATTACATCA	3.76 ± 0.77	8.96 ± 0.52	0.16	7

Nucleotide sequence	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	Reference
	Amber99sb*ILDN-BSC1	Charmm36m	Experimental	
CATGTAAT G AATTACATCA	10.16 ± 1.23	9.39 ± 1.15	2.00	7
CATGTAATT C ATTACATCA	4.82 ± 1.09	10.89 ± 0.97	2.82	7
CATGTAATT T GATTACATCA	7.26 ± 0.75	10.31 ± 0.89	1.68	7
CATGTAATT T TATTACATCA	1.84 ± 1.31	3.69 ± 1.02	0.32	7
CATGTAATT A CTTACATCA	-1.49 ± 0.92	-1.42 ± 1.85	1.14	7
CATGTAATT A GTTACATCA	1.94 ± 0.59	2.64 ± 0.82	0.91	7
CATGTAATT A A A TACATCA	0.57 ± 1.20	-1.36 ± 0.84	-1.04	7
CATGTAATT A A G TACATCA	2.97 ± 1.36	-7.04 ± 1.14	-1.68	7
CATGTAATT A A A A TACATCA	-1.45 ± 3.31	-3.94 ± 1.30	0.32	7
CATGTAATT A A A T CACATCA	4.87 ± 0.79	-3.69 ± 0.69	-0.36	7
CATGTAATT A A A T G ACATCA	13.29 ± 1.34	-7.74 ± 1.26	5.58	7
CATGTAATT A A A T T CCATCA	-1.01 ± 1.83	4.03 ± 1.16	0.16	7
CATGTAATT A A A T T G CATCA	-0.74 ± 0.70	5.51 ± 0.53	0.32	7
CATGTAATT A A A T T T CATCA	13.22 ± 1.94	21.10 ± 1.81	5.58	7
CATGTAATT A A A T T A ATCA	9.57 ± 1.25	10.39 ± 1.22	0.32	7
CATGTAATT A A A T T A T GATCA	14.07 ± 1.72	10.80 ± 1.11	4.81	7
CATGTAATT A A A T T A T A TATCA	4.02 ± 1.04	10.84 ± 0.94	1.68	7
CATGTAATT A A A T T A T T ACTCA	7.52 ± 1.89	16.26 ± 1.01	0.32	7
CATGTAATT A A A T T A T T A CGTCA	8.72 ± 1.47	12.26 ± 0.79	6.72	7
CATGTAATT A A A T T A T T A T CTCA	15.06 ± 1.53	10.44 ± 1.56	1.39	7
CATGTAATT A A A T T A T T A T A ACA	12.71 ± 1.44	12.48 ± 1.75	5.91	7
CATGTAATT A A A T T A T T A T A ACCCA	6.58 ± 0.88	6.66 ± 0.66	4.06	7
CATGTAATT A A A T T A T T A T A AGCA	14.31 ± 1.41	18.32 ± 1.03	6.28	7
CATGTAATT A A A T T A T T A T A ATA	7.35 ± 1.78	5.97 ± 1.54	2.38	7
CATGTAATT A A A T T A T T A T G A	17.58 ± 1.62	21.23 ± 1.13	2.54	7
CATGTAATT A A A T T A T T A T T TA	13.70 ± 1.41	13.51 ± 1.26	0.32	7
CATGTAATT A A A T T A T T A T C CC	-0.90 ± 1.38	2.77 ± 1.17	-1.18	7
CATGTAATT A A A T T A T T A T C T	2.40 ± 1.51	4.35 ± 1.77	-1.52	7

BamHI

ATGGATCCATA				
T GGGATCCC C	-23.28 ± 4.07	-2.14 ± 6.09	-0.42	8
C GGGATCCC G	-7.03 ± 5.09	-2.17 ± 5.81	0.84	8
G GGGATCC G C	-17.31 ± 3.19	8.40 ± 4.09	0.84	8
T AGGATC C T A	0.85 ± 6.75	24.58 ± 5.82	0.84	8
ACGGATCC G T	-8.64 ± 1.61	10.23 ± 1.89	0.84	8
CTGGATCC A G C	-23.27 ± 5.48	-12.53 ± 3.97	1.26	8
C CGGATCC G G G	-23.56 ± 4.96	15.50 ± 5.64	1.26	8
G TGGATCC A C C	-21.56 ± 5.93	-3.24 ± 2.72	1.67	8

Nucleotide sequence	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	Reference
	Amber99sb*ILDN-BSC1	Charmm36m	Experimental	
GTGGATCCACA	-7.03 ± 2.29	-9.76 ± 0.98	1.67	8
GCGGATCCGCG	3.97 ± 8.09	11.74 ± 3.23	1.67	8
AAGGATCCTTA	13.67 ± 5.33	13.32 ± 3.97	2.09	8
TAGGATCCTAA	-1.95 ± 4.25	17.13 ± 4.10	2.93	8
GTGGATCCACG	-11.43 ± 2.76	3.74 ± 1.52	2.93	8
TTGGATCCAAG	-7.23 ± 4.02	2.03 ± 3.32	3.35	8
CAGGATCCTGG	-16.68 ± 5.77	8.78 ± 6.66	3.35	8
AGGGATCCCTT	13.01 ± 7.90	3.77 ± 5.16	4.60	8
AAGGATCCTTG	19.55 ± 4.69	13.45 ± 2.19	4.60	8
GGGGATCCCCC	-10.66 ± 5.58	-18.05 ± 7.10	5.02	8
AAGGATCCTTT	20.07 ± 5.40	0.96 ± 3.30	5.02	8
GAGGATCCTCT	15.85 ± 4.39	5.12 ± 4.69	5.44	8
GAGGATCCTCG	8.67 ± 7.97	15.93 ± 2.91	5.86	8
GGGGATCCCCA	-0.85 ± 4.13	-2.09 ± 5.80	6.69	8
GGGGATCCCCG	-14.57 ± 4.81	13.59 ± 5.97	7.95	8
Tus-Ter				
AGTATGTTGTAACT				
AATATGTTGTAACT	-2.91 ± 0.97	-0.54 ± 0.57	3.47	9
ACTATGTTGTAACT	4.43 ± 1.40	1.04 ± 1.02	1.51	9
ATTATGTTGTAACT	-5.31 ± 1.40	-3.37 ± 0.94	3.18	9
AGAATGTTGTAACT	6.42 ± 1.39	0.09 ± 1.19	4.14	9
AGGATGTTGTAACT	-0.24 ± 1.43	-0.10 ± 0.94	1.00	9
AGTCTGTTGTAACT	14.38 ± 1.80	12.54 ± 0.93	10.75	9
AGTGTGTTGTAACT	3.54 ± 0.89	2.74 ± 0.55	6.90	9
AGTTTGTTGTAACT	6.00 ± 1.95	7.80 ± 1.12	11.67	9
AGTAAGTTGTAACT	10.09 ± 1.65	7.66 ± 1.87	8.16	9
AGTACGTTGTAACT	1.41 ± 0.92	6.72 ± 1.09	7.91	9
AGTAGGTTGTAACT	12.33 ± 1.76	8.44 ± 0.88	9.83	9
AGTATAATTGTAACT	12.92 ± 1.02	12.93 ± 1.27	16.74*	9
AGTATCTTGTAACT	10.06 ± 2.49	16.74 ± 1.48	16.74*	9
AGTATTTGTAACT	14.65 ± 2.61	16.74 ± 1.33	16.74*	9
AGTATGATGTAACT	12.03 ± 1.33	3.77 ± 1.55	9.67	9
AGTATGCTGTAACT	8.49 ± 0.96	14.03 ± 1.36	10.08	9
AGTATGGTGTAACT	18.23 ± 1.54	14.49 ± 1.79	12.22	9
AGTATGTAGTAACT	16.74 ± 3.08	16.74 ± 2.82	16.74*	9
AGTATGTCGTAACT	10.50 ± 1.60	8.08 ± 0.99	16.74*	9
AGTATGTGGTAACT	11.50 ± 3.10	7.18 ± 2.30	16.74*	9
AGTATGTTATAACT	16.74 ± 2.20	7.47 ± 1.84	16.74*	9

Nucleotide sequence	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	Reference
	Amber99sb*ILDN-BSC1	Charmm36m	Experimental	
AGTATGTTCTAACT	0.22 ± 2.54	16.16 ± 1.27	16.74*	9
AGTATGTTTTAACT	9.73 ± 3.35	-3.26 ± 1.50	16.74*	9
AGTATGTTGAAACT	16.74 ± 2.97	3.31 ± 2.21	16.74*	9
AGTATGTTGCAACT	16.74 ± 1.15	-4.64 ± 4.31	16.74*	9
AGTATGTTGGAACT	10.62 ± 1.79	-9.74 ± 3.08	16.74*	9
AGTATGTTGTCACT	0.87 ± 1.63	11.53 ± 2.23	12.97	9
AGTATGTTGTGACT	15.79 ± 0.90	11.86 ± 1.68	16.74*	9
AGTATGTTGTTACT	11.17 ± 1.41	0.97 ± 2.59	16.74*	9
AGTATGTTGTACCT	8.41 ± 1.39	-5.44 ± 1.97	16.61	9
AGTATGTTGTAGCT	21.71 ± 1.01	4.37 ± 1.03	16.23	9
AGTATGTTGTATCT	5.88 ± 2.18	-2.70 ± 3.59	16.74*	9
AGTATGTTGTAAAT	9.90 ± 1.87	11.60 ± 1.23	16.74*	9
AGTATGTTGTAAGT	8.81 ± 1.69	11.69 ± 2.27	16.74*	9
AGTATGTTGTAATT	9.41 ± 1.36	2.92 ± 0.74	7.15	9
AGTATGTTGTAACA	-4.10 ± 1.77	0.54 ± 1.74	16.74*	9
AGTATGTTGTAACC	-2.24 ± 0.69	-3.47 ± 0.65	15.52	9
AGTATGTTGTAACG	-0.32 ± 2.20	1.50 ± 2.27	10.25	9
LacR				
TGTGAGCGCTCACATT				
TGTAAGCGCTTACAATT	-30.88 ± 4.39	10.54 ± 4.83	16.74	10
TGAGAGCGCTCTCAATT	14.35 ± 3.22	0.40 ± 3.37	13.81	10
TGCGAGCGCTCGCAATT	7.08 ± 4.62	21.43 ± 1.30	20.50	10
TGAAAGCGCTTCAATT	-11.69 ± 5.22	17.13 ± 4.87	21.34	10
TGCAAGCGCTTGCAATT	9.92 ± 3.98	39.22 ± 4.19	28.45	10
ER				
GAGGTCACAGTGACCTG				
GTGGTCACAGTGACCAAG	12.10 ± 2.58	2.75 ± 2.21	15.40	11
GACGTCACAGTGACGTG	41.32 ± 2.34	55.90 ± 3.70	17.88	11
GAGATCACAGTGATCTG	18.85 ± 2.70	39.39 ± 1.69	17.14	11
GAGGACACAGTGCCTG	31.77 ± 4.84	20.65 ± 2.30	16.87	11
GAGGTGACAGTCACCTG	24.82 ± 2.67	57.95 ± 2.47	21.02	11
GAGGTCTCAGAGACCTG	2.99 ± 4.39	8.53 ± 2.42	16.42	11
GGGGCCGGGCTGACCCG	4.33 ± 4.20	1.95 ± 3.14	13.55	11
CroR				
CTATCACCGCCAGAGGTAC				
CAATCACCGCCAGAGGTAC	8.18 ± 1.36	3.05 ± 1.24	4.60	12

Nucleotide sequence	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	Reference
	Amber99sb*ILDN-BSC1	Charmm36m	Experimental	
CGATCACCGCCAGAGGTAC	2.97 ± 1.44	7.90 ± 1.24	6.28	12
CCATCACCGCCAGAGGTAC	3.10 ± 0.80	2.68 ± 0.91	5.44	12
CT G TACCCGCCAGAGGTAC	13.61 ± 0.90	15.71 ± 0.56	11.72	12
CT T TCACCGCCAGAGGTAC	-11.85 ± 2.20	13.49 ± 3.19	10.46	12
CT C TCACCGCCAGAGGTAC	2.74 ± 1.37	15.02 ± 1.01	12.97	12
CT A GCACCGCCAGAGGTAC	11.96 ± 1.22	11.84 ± 1.74	6.28	12
CT A ACACCGCCAGAGGTAC	4.98 ± 1.98	1.91 ± 2.40	2.51	12
CT A CCACCGCCAGAGGTAC	12.83 ± 1.01	8.24 ± 1.18	7.11	12
CT A TGACCGCCAGAGGTAC	3.43 ± 2.20	-1.66 ± 1.79	11.30	12
CT A TAACCGCCAGAGGTAC	2.50 ± 4.18	12.99 ± 1.47	10.04	12
CT A TATTACCGCCAGAGGTAC	1.82 ± 0.69	-0.18 ± 0.54	5.86	12
CT A TATCCCCGCCAGAGGTAC	1.20 ± 2.21	6.63 ± 2.08	6.69	12
CT A TATCTCCGCCAGAGGTAC	11.09 ± 1.23	8.74 ± 1.66	5.02	12
CT A TATGCCGCCAGAGGTAC	6.79 ± 1.23	-0.08 ± 1.13	7.11	12
CT A TATCAGGCCAGAGGTAC	7.72 ± 1.19	13.26 ± 1.56	9.20	12
CT A TATCAACGCCAGAGGTAC	9.04 ± 2.01	8.26 ± 1.70	7.53	12
CT A TATCATGCCAGAGGTAC	4.74 ± 0.85	4.20 ± 0.66	9.62	12
CT A TACTACTGCCAGAGGTAC	-0.63 ± 0.73	2.97 ± 0.67	1.67	12
CT A TATCACGGCCAGAGGTAC	2.34 ± 1.19	4.29 ± 0.88	8.37	12
CT A TATCACAGGCCAGAGGTAC	3.79 ± 1.39	3.81 ± 0.93	10.88	12
CT A TATCACCCCCAGAGGTAC	4.85 ± 1.07	0.75 ± 1.26	0.21	12
CT A TATCACCTCCAGAGGTAC	-0.88 ± 1.41	-4.57 ± 1.03	-0.84	12
CT A TATCACCAACCAGAGGTAC	3.49 ± 0.75	-2.39 ± 0.63	-0.42	12
CT A TATCACCGGCAGAGGTAC	1.02 ± 1.11	-1.00 ± 0.90	0.42	12
CT A TATCACCGTCAGAGGTAC	4.58 ± 0.63	-5.22 ± 0.71	2.51	12
CT A TATCACCGACAGAGGTAC	4.63 ± 1.04	-3.73 ± 1.06	2.72	12
CT A TATCACCGCTAGAGGTAC	-2.72 ± 0.95	-3.30 ± 0.66	-0.84	12
CT A TATCACCGCAGAGGTAC	0.99 ± 2.50	-0.86 ± 0.83	-1.26	12
CT A TATCACCGCGAGAGGTAC	8.89 ± 1.99	0.72 ± 1.07	0.00	12
CT A TATCACCGCCTGAGAGGTAC	-7.23 ± 1.49	5.02 ± 1.19	7.95	12
CT A TATCACCGCCCGAGAGGTAC	-0.64 ± 1.09	0.97 ± 1.03	5.44	12
CT A TATCACCGCCGGAGAGGTAC	0.29 ± 0.71	-0.28 ± 0.57	-1.67	12
CT A TATCACCGCCAAGAGGTAC	9.10 ± 0.75	6.50 ± 0.60	9.00	12
CT A TATCACCGCCATAGGTAC	15.96 ± 1.58	11.18 ± 1.75	7.53	12
CT A TATCACCGCCACAGAGGTAC	10.32 ± 1.08	12.98 ± 1.70	9.00	12
CT A TATCACCGCCAGCGGTAC	-4.20 ± 1.95	-5.34 ± 1.15	4.18	12
CT A TATCACCGCCAGTGGTAC	-7.73 ± 2.37	-5.03 ± 1.00	-2.51	12
CT A TATCACCGCCAGGGGTAC	-12.49 ± 1.69	-1.52 ± 1.39	3.35	12
CT A TATCACCGCCAGAAGTAC	9.37 ± 1.06	7.56 ± 0.74	7.11	12

Nucleotide sequence	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	Reference
	Amber99sb*ILDN-BSC1	Charmm36m	Experimental	
CTATCACCGCCAGATGTAC	1.73 ± 2.87	4.78 ± 1.13	7.95	12
CTATCACCGCCAGACGTAC	-0.93 ± 2.44	3.59 ± 1.33	4.18	12
CTATCACCGCCAGAGATAAC	-10.13 ± 1.15	-10.36 ± 0.75	-5.44	12
CTATCACCGCCAGAGTTAC	-6.85 ± 1.23	-4.11 ± 1.19	-2.93	12
CTATCACCGCCAGAGCTAC	-3.26 ± 1.57	1.70 ± 1.25	1.67	12
CTATCACCGCCAGAGGGAC	2.34 ± 1.46	5.62 ± 1.03	7.74	12
CTATCACCGCCAGAGGAAC	3.01 ± 3.54	11.32 ± 1.72	7.53	12
CTATCACCGCCAGAGGCAC	4.52 ± 1.04	4.62 ± 0.84	6.90	12
CTATCACCGCCAGAGGTGC	0.53 ± 0.63	1.79 ± 0.64	3.14	12
CTATCACCGCCAGAGGTCC	1.66 ± 1.25	-0.14 ± 1.21	2.72	12
CTATCACCGCCAGAGGTTTC	0.91 ± 1.29	-2.93 ± 1.15	0.63	12
CTAACACCGTGCAGAGTTGC	22.07 ± 3.82	-8.48 ± 5.61	5.44	12
CTATCACCGCAAGGGATAC	-8.79 ± 5.51	-7.47 ± 1.93	-4.60	12
CTATCACCGCCAGTGGTAC	-10.63 ± 1.58	-1.11 ± 1.05	-0.42	12
CCAACACCGCCAGAGATAAC	-0.45 ± 3.65	5.06 ± 3.66	2.09	12
CTATCACCGCAGATGGTTC	-6.39 ± 5.74	-7.85 ± 6.52	4.18	12
EcoRI				
GCAGAATTCTGC				
GCATAATTCTGC	31.65 ± 4.27	-9.06 ± 3.44	17.15	13
GCAAAATTCTGC	7.19 ± 0.76	10.93 ± 1.48	18.41	13
GCACAATTCTGC	21.85 ± 1.67	8.94 ± 1.47	21.76	13
GCAGCATTCTGC	17.39 ± 1.69	26.37 ± 2.18	22.18	13
GCAGGATTCTGC	20.89 ± 1.07	22.26 ± 1.48	23.43	13
GCAGTATTCTGC	15.51 ± 2.22	21.12 ± 2.33	18.83	13
GCAGACTTCTGC	11.24 ± 1.72	14.08 ± 1.45	21.34	13
GCAGAGTTCTGC	0.06 ± 1.18	13.25 ± 2.35	19.66	13
GCAGATTCTGC	5.32 ± 1.75	18.52 ± 3.74	22.59	13
GCAGGATCCTGC	23.84 ± 2.01	27.10 ± 2.22	20.08	13
GCAGAGCTCTGC	2.44 ± 1.84	34.76 ± 1.69	22.18	13
GCAAATTATGC	25.39 ± 2.85	26.82 ± 5.06	23.85	13
GCACCTAAGTGC	32.69 ± 12.91	75.82 ± 8.63	21.76	13
CAP				
CGAAAAATGTGATCTAGATCACATTTTCG				
CGAAAAAAAGTGATCTAGATCACTTTTCG	1.18 ± 3.59	4.73 ± 2.15	10.46	14
CGAAAAAACGTGATCTAGATCACGTTTTCG	3.72 ± 1.55	7.12 ± 0.99	3.77	14
CGAAAAAAAGGTGATCTAGATCACCTTTTCG	-7.89 ± 2.48	0.85 ± 4.67	9.62	14
CGAAAAAAATGTGATCTAGATCATTTTTCG	9.60 ± 0.93	15.19 ± 1.69	19.25	14

Nucleotide sequence	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	Reference
	Amber99sb*ILDN-BSC1	Charmm36m	Experimental	
CGAAAAAATCTGATCTAGATCAGATTTTCG	20.96 ± 4.71	21.76 ± 2.85	21.76*	14
CGAAAAAATTGATCTAGATCAAATTTTCG	22.11 ± 1.82	27.97 ± 2.61	12.55	14
CGAAAATGAGATCTAGATCTCATTTTCG	8.40 ± 4.60	15.62 ± 1.71	12.13	14
CGAAAATGCGATCTAGATCGCATTTCG	10.97 ± 2.84	24.33 ± 1.75	11.72	14
CGAAAATGGGATCTAGATCCATTTTCG	17.77 ± 5.00	13.56 ± 2.00	11.72	14
CGAAAATGTAACTAGATTACATTTTCG	21.76 ± 3.66	21.76 ± 1.58	21.76*	14
CGAAAATGTCATCTAGATGACATTTTCG	21.76 ± 2.78	21.76 ± 5.35	21.76*	14
CGAAAATGTTATCTAGATAACATTTTCG	23.05 ± 3.00	24.75 ± 4.35	19.66	14
CGAAAATGTGCTCTAGAGCACATTTTCG	-7.11 ± 3.00	5.94 ± 2.26	9.62	14
CGAAAATGTGGCTAGACCACATTTTCG	3.17 ± 2.60	2.67 ± 2.97	14.23	14
CGAAAATGTGTTCTAGAACACATTTTCG	18.03 ± 2.62	-0.45 ± 2.23	12.97	14

PU.1 ETS

ATAAGGGAAAGTGAA				
ATAAAAGGAAGTGAA	2.91 ± 1.47	-0.55 ± 1.05	0.71	15
ATACAAGGAAGTGAA	8.56 ± 2.93	0.85 ± 2.18	5.19	15
ATAGAAGGAAGTGAA	4.76 ± 1.94	2.62 ± 1.82	6.11	15
ATATAAGGAAGTGAA	9.65 ± 2.41	3.50 ± 1.80	1.09	15
ATAACAGGAAGTGAA	-4.07 ± 2.62	-0.90 ± 1.41	-0.40	15
ATAAGAGGAAGTGAA	4.67 ± 0.91	5.94 ± 0.74	-2.85	15
ATAAACCGGAAGTGAA	-0.88 ± 1.71	2.34 ± 1.54	4.27	15
ATAAAGGGAAAGTGAA	1.10 ± 0.79	-3.83 ± 0.62	6.02	15
ATAAATGGAAGTGAA	14.25 ± 1.98	15.04 ± 1.38	10.46	15
ATAAAAGGAAATGAA	10.70 ± 2.94	-6.33 ± 1.00	9.00	15
ATAAAAGGAAGAGAA	9.18 ± 5.58	-3.27 ± 2.32	6.74	15
ATAAAAGGAATTGAA	12.52 ± 3.65	-4.70 ± 1.86	10.54	15
ATAAAAGGAAGGGAA	14.57 ± 2.18	-2.32 ± 1.46	9.37	15
ATAACCGGAAGTGAA	-1.91 ± 2.76	3.72 ± 1.82	-1.76	15
ATAACCGGAAGTGAA	2.96 ± 1.74	-1.10 ± 0.92	7.36	15
ATAACTGGAAGTGAA	6.61 ± 1.78	7.37 ± 3.13	7.70	15
ATAAGCGGAAGTGAA	-2.30 ± 1.67	1.27 ± 1.22	-3.31	15
ATAAGTGGAAAGTGAA	11.96 ± 1.31	12.62 ± 1.24	5.36	15
ATATGAGGAAGTGAA	14.70 ± 4.50	10.42 ± 1.50	1.13	15
ATATACCGGAAGTGAA	0.14 ± 2.50	11.67 ± 2.74	2.51	15
ATAAAAGGAAAAGAA	5.74 ± 2.73	0.73 ± 2.55	9.08	15
ATAAAAGGAATGGAA	22.67 ± 3.53	-2.99 ± 2.79	11.84	15
ATATGCCGAAGTGAA	2.84 ± 3.99	7.87 ± 2.03	1.30	15
ATAAGAGGAAGGGAA	11.96 ± 3.42	7.15 ± 1.51	8.16	15
ATAAAACGGAAATGAA	4.29 ± 2.79	2.69 ± 2.01	9.33	15

Nucleotide sequence	$\Delta\Delta G$, kJ/mol Amber99sb*ILDN-BSC1	$\Delta\Delta G$, kJ/mol Charmm36m	$\Delta\Delta G$, kJ/mol Experimental	Reference
Ndt80				
GTGTCACAAATT _A				
T TGTCACAAATT _A	-0.01 ± 1.50	3.77 ± 1.49	1.68	16
C TGTCACAAATT _A	2.35 ± 1.86	-3.78 ± 1.85	1.28	16
A TGTCACAAATT _A	0.99 ± 0.73	1.61 ± 0.47	0.83	16
G AGTCACAAATT _A	-7.46 ± 1.49	0.34 ± 1.21	0.45	16
G GGTCACAAATT _A	-6.13 ± 1.50	5.14 ± 1.05	2.22	16
G CGTCACAAATT _A	-7.16 ± 0.87	-1.22 ± 0.54	1.12	16
G TATCACAAATT _A	8.76 ± 0.94	19.55 ± 0.60	-0.25	16
G TCTCACAAATT _A	2.39 ± 3.47	28.04 ± 1.62	3.09	16
G TTCACAAATT _A	11.99 ± 2.38	25.15 ± 1.90	1.41	16
G TGCCACAAATT _A	0.15 ± 0.78	0.39 ± 0.57	-0.87	16
G TGACACAAATT _A	-2.47 ± 1.86	2.59 ± 1.22	-1.68	16
G TGGCACAAATT _A	-7.22 ± 1.40	-0.48 ± 1.10	0.00	16
G TGTTACAAATT _A	10.12 ± 1.12	31.21 ± 0.77	3.46	16
G TGTAACAAATT _A	3.77 ± 2.83	26.22 ± 0.88	6.12	16
G TGTCGCAAATT _A	1.08 ± 1.02	5.62 ± 0.61	3.00	16
G TGTCCCCAAATT _A	-5.22 ± 1.46	-1.85 ± 0.95	4.30	16
G TGTCATAAATT _A	8.17 ± 1.14	18.20 ± 0.65	9.48	16
G TGTCAAAATT _A	47.41 ± 2.49	28.00 ± 1.20	9.48	16
G TGTCACTAATT _A	-1.99 ± 1.53	-2.51 ± 1.05	5.14	16
G TGTCACGAATT _A	5.64 ± 0.98	4.25 ± 0.63	4.95	16
G TGTCACATATT _A	3.76 ± 1.59	0.12 ± 1.04	4.16	16
G TGTCACAGATT _A	4.36 ± 0.68	-5.80 ± 0.53	4.77	16
G TGTCACAATT _A	8.30 ± 1.12	0.45 ± 1.12	2.84	16
G TGTCACAAGTT _A	6.04 ± 0.88	9.36 ± 0.67	2.10	16
G TGTCACAAAGTA	-5.66 ± 1.18	3.73 ± 1.24	-1.23	16
G TGTCACAAAT G A	3.02 ± 1.15	-0.61 ± 0.96	0.00	16
c-Myb				
CCTAACTGACA				
C CGAACTGACA	-8.46 ± 1.80	3.41 ± 2.42	3.75	17,18
C CAAAC TGACA	-3.53 ± 1.52	-1.15 ± 3.19	2.42	17
C CCAAC TGACA	-3.82 ± 1.12	-1.42 ± 1.43	0.35	17
C CT G ACTGACA	6.38 ± 1.06	-0.77 ± 0.75	9.23	18–20
C CTTACTGACA	-13.30 ± 3.77	-0.38 ± 1.31	11.30	19
C CT C ACTGACA	-5.21 ± 1.61	2.49 ± 1.47	12.13	19
C CTAGCTGACA	2.77 ± 1.24	2.06 ± 0.76	7.26	18–20

Nucleotide sequence	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	$\Delta\Delta G$, kJ/mol	Reference
	Amber99sb*ILDN-BSC1	Charmm36m	Experimental	
CCTAT T CTGACA	1.89 ± 2.55	12.20 ± 2.91	6.69	18,19
CCTAC CT TGACA	-1.54 ± 1.56	-8.38 ± 1.04	4.81	18,19
CCTAAG T GACA	6.39 ± 1.46	6.31 ± 1.80	12.97	19
CCTAA A TGACA	0.76 ± 1.90	1.92 ± 2.15	12.55	19
CCTAAT T GACA	7.81 ± 1.55	4.01 ± 0.99	10.20	18–20
CCTAAC CG GACA	9.03 ± 2.08	7.10 ± 4.46	1.26	19
CCTAAC A GACA	14.37 ± 1.84	12.59 ± 2.29	4.60	19
CCTAAC CC GACA	-1.71 ± 1.39	6.65 ± 0.93	0.00	19
CCTAA CT AACA	20.42 ± 1.36	13.88 ± 1.19	7.52	18–20
CCTAA CTT AACA	19.48 ± 1.76	17.52 ± 2.29	7.95	18,19
CCTAA ACT CACA	26.92 ± 1.44	31.69 ± 2.58	13.39	19
CCTAA ACTG CA	-9.88 ± 1.49	-11.26 ± 0.90	2.51	19
CCTAA ACTGT CA	-3.58 ± 1.68	-16.52 ± 1.78	-0.84	19
CCTAA ACTGCC A	3.28 ± 2.20	-8.11 ± 1.97	-1.67	19
CCTAA ACTGAG A	7.73 ± 1.32	1.86 ± 0.97	7.95	18,19
CCTAA ACTGAA A	2.99 ± 1.61	5.75 ± 1.32	2.51	19
CCTAA ACTGATA	2.18 ± 1.01	-0.00 ± 0.50	3.14	18,19
CCTAA ACTGAC G	-1.38 ± 0.77	-2.44 ± 0.44	1.42	21
CCTAA ACTGACT	-1.40 ± 2.08	0.53 ± 2.59	0.17	21
CCTAA ACTGACC	-3.77 ± 1.89	1.17 ± 1.51	1.84	21

* A lower bound for the $\Delta\Delta G$ value is provided by an experiment. If a calculated value exceeds the experimental lower bound, the calculated $\Delta\Delta G$ is capped at the experimental estimate.

Experimental values with several references were combined by taking an average $\Delta\Delta G$ of the values reported in the literature sources.

References

- (1) Hamilton, T. B.; Borel, F.; Romaniuk, P. J. Comparison of the DNA binding characteristics of the related zinc finger proteins WT1 and EGR1. *Biochemistry* **1998**, *37*, 2051–2058.
- (2) Miller, J. C.; Pabo, C. O. Rearrangement of side-chains in a Zif268 mutant highlights the complexities of zinc finger-DNA recognition. *J. Mol. Biol.* **2001**, *313*, 309–315.
- (3) Sarai, A.; Takeda, Y. Lambda repressor recognizes the approximately 2-fold symmetric half-operator sequences asymmetrically. *Proc. Natl. Acad. Sci. U. S. A.* **1989**, *86*, 6513–6517.
- (4) Grillo, A. O.; Brown, M. P.; Royer, C. A. Probing the physical basis for trp repressor-operator recognition. *J. Mol. Biol.* **1999**, *287*, 539–554.
- (5) Hao, D.; Ohme-Takagi, M.; Sarai, A. Unique mode of GCC box recognition by the DNA-binding domain of ethylene-responsive element-binding factor (ERF domain) in plant. *J. Biol. Chem.* **1998**, *273*, 26857–26861.
- (6) Fujimoto, S. Y.; Ohta, M.; Usui, A.; Shinshi, H.; Ohme-Takagi, M. Arabidopsis ethylene-responsive element binding factors act as transcriptional activators or repressors of GCC box-mediated gene expression. *The Plant Cell* **2000**, *12*, 393–404.
- (7) Jin, Y.; Zhong, H.; Vershon, A. K. The yeast α 1 and α 2 homeodomain proteins do not contribute equally to heterodimeric DNA binding. *Mol. Cell. Biol.* **1999**, *19*, 585–593.
- (8) Engler, L. E.; Sapienza, P.; Dorner, L. F.; Kucera, R.; Schildkraut, I.; Jen-Jacobson, L. The energetics of the interaction of BamHI endonuclease with its recognition site GGATCC. *J. Mol. Biol.* **2001**, *307*, 619–636.
- (9) Coskun-Ari, F. F.; Hill, T. M. Sequence-specific interactions in the Tus-Ter complex

- and the effect of base pair substitutions on arrest of DNA replication in *Escherichia coli*. *J. Biol. Chem.* **1997**, *272*, 26448–26456.
- (10) Frank, D. E.; Saecker, R. M.; Bond, J. P.; Capp, M. W.; Tsodikov, O. V.; Melcher, S. E.; Levandoski, M. M.; Record, M. T. Thermodynamics of the interactions of lac repressor with variants of the symmetric lac operator: effects of converting a consensus site to a non-specific site. *J. Mol. Biol.* **1997**, *267*, 1186–1206.
- (11) Boyer, M.; Poujol, N.; Margeat, E.; Royer, C. A. Quantitative characterization of the interaction between purified human estrogen receptor α and DNA using fluorescence anisotropy. *Nucleic Acids Res.* **2000**, *28*, 2494–2502.
- (12) Takeda, Y.; Sarai, A.; Rivera, V. M. Analysis of the sequence-specific interactions between Cro repressor and operator DNA by systematic base substitution experiments. *Proc. Natl. Acad. Sci. U. S. A.* **1989**, *86*, 439–443.
- (13) Lesser, D. R.; Kurpiewski, M. R.; Jen-Jacobson, L. The energetic basis of specificity in the EcoRI endonuclease-DNA interaction. *Science* **1990**, *250*, 776–786.
- (14) Gunasekera, A.; Ebright, Y. W.; Ebright, R. H. DNA sequence determinants for binding of the *Escherichia coli* catabolite gene activator protein. *J. Biol. Chem.* **1992**, *267*, 14713–14720.
- (15) Poon, G. M. K.; Macgregor Jr, R. B. Base coupling in sequence-specific site recognition by the ETS domain of murine PU.1. *J. Mol. Biol.* **2003**, *328*, 805–819.
- (16) Pierce, M.; Benjamin, K. R.; Montano, S. P.; Georgiadis, M. M.; Winter, E.; Vershon, A. K. Sum1 and Ndt80 proteins compete for binding to middle sporulation element sequences that control meiotic gene expression. *Mol. Cell. Biol.* **2003**, *23*, 4814–4825.

- (17) Oda, M.; Furukawa, K.; Ogata, K.; Sarai, A.; Ishii, S.; Nishimura, Y.; Nakamura, H. Investigation of the pyrimidine preference by the c-Myb DNA-binding domain at the initial base of the consensus sequence. *J. Biol. Chem.* **1997**, *272*, 17966–17971.
- (18) Oda, M.; Furukawa, K.; Ogata, K.; Sarai, A.; Nakamura, H. Thermodynamics of specific and non-specific DNA binding by the c-Myb DNA-binding domain. *J. Mol. Biol.* **1998**, *276*, 571–590.
- (19) Ogata, K.; Kanei-Ishii, C.; Sasaki, M.; Hatanaka, H.; Nagadoi, A.; Enari, M.; Nakamura, H.; Nishimura, Y.; Ishii, S.; Sarai, A. The cavity in the hydrophobic core of Myb DNA-binding domain is reserved for DNA recognition and trans-activation. *Nat. Struct. Mol. Biol.* **1996**, *3*, 178–187.
- (20) Oda, M.; Furukawa, K.; Sarai, A.; Nakamura, H. Kinetic analysis of DNA binding by the c-Myb DNA-binding domain using surface plasmon resonance. *FEBS Lett.* **1999**, *454*, 288–292.
- (21) Tanikawa, J.; Yasukawa, T.; Enari, M.; Ogata, K.; Nishimura, Y.; Ishii, S.; Sarai, A. Recognition of specific DNA sequences by the c-myb protooncogene product: role of three repeat units in the DNA-binding domain. *Proc. Natl. Acad. Sci. U. S. A.* **1993**, *90*, 9320–9324.